

*RT with CAR-T:  
Bridging, Priming, Debulking,  
Consolidation or **All the Above***

***Bouthaina Dabaja, MD***  
***University of Texas MD Anderson Cancer Center***

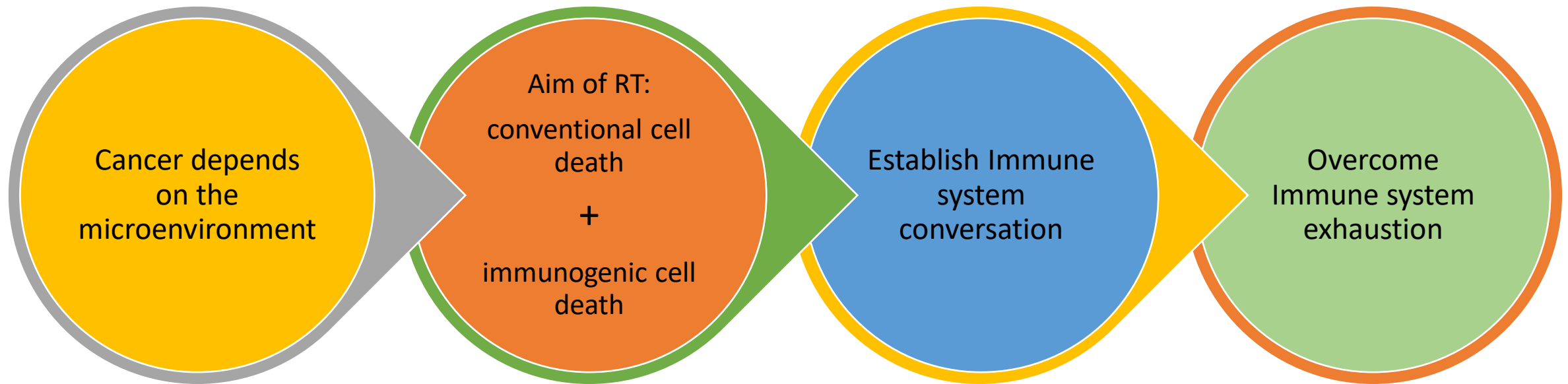


# Disclosures

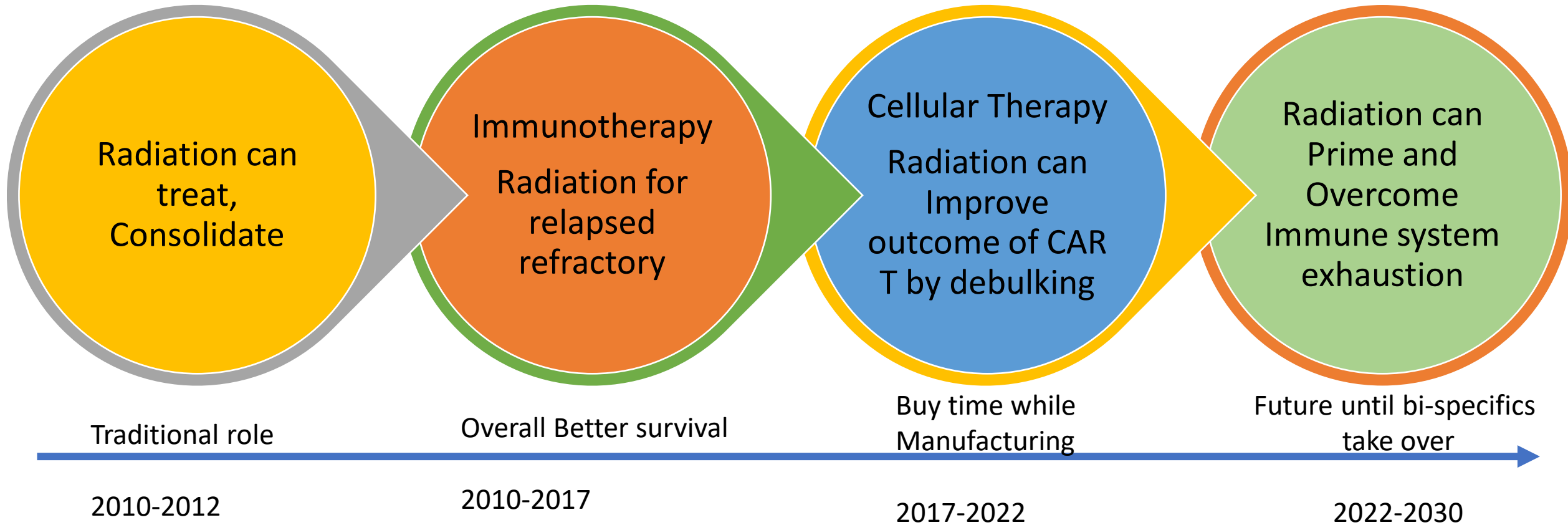
## Research supports

- Seattle Genetics
- Kite
  
- Advisory to ONO-Pharma

# The new paradigm of Cancer Treatment



# Timeline of RT role



**Conduct translation studies on the effect/ timing of radiation rather than a binary response / no response**

# Objectives of my talk



Landscape of Cellular and Immunotherapy



Early evidence for the clinical benefit of radiation



What we know about priming with radiation



The current ongoing research and future direction



# FDA-Approved CAR T-Cell Therapies

Generic Name	Brand Name	Target Antigen	Targeted Disease	Patient Population
Tisagenlecleucel	Kymriah	CD19	B-cell acute lymphoblastic leukemia (ALL)	Children and young adults with refractory or relapsed B-cell ALL
			B-cell non-Hodgkin lymphoma (NHL)	Adults with relapsed or refractory B-cell NHL
Axicabtagene ciloleucel	Yescarta	CD19	B-cell non-Hodgkin lymphoma (NHL)	Adults with relapsed or refractory B-cell NHL
			Follicular lymphoma	Adults with relapsed or refractory follicular lymphoma
Brexucabtagene autoleucel	Tecartus	CD19	Mantle cell lymphoma (MCL)	Adults with relapsed or refractory MCL
			B-cell acute lymphoblastic leukemia (ALL)	Adults with refractory or relapsed B-cell ALL
Lisocabtagene maraleucel	Breyanzi	CD19	B-cell non-Hodgkin lymphoma (NHL)	Adults with relapsed or refractory B-cell NHL
Idecabtagene vicleucel	Abecma	BCMA	Multiple myeloma	Adults with relapsed or refractory multiple myeloma
Ciltacabtagene autoleucel	Carvykti	BCMA	Multiple myeloma	Adults with relapsed or refractory multiple myeloma

CAR structure

BBz, lenti

28z,  $\gamma$ -retro

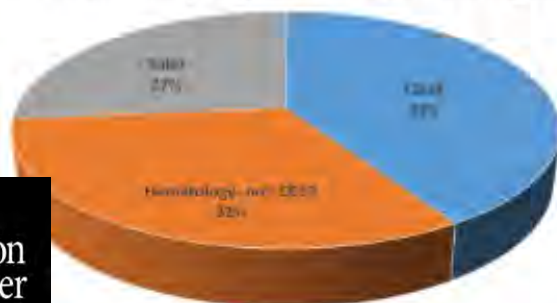
28z,  $\gamma$ -retro

BBz, lenti

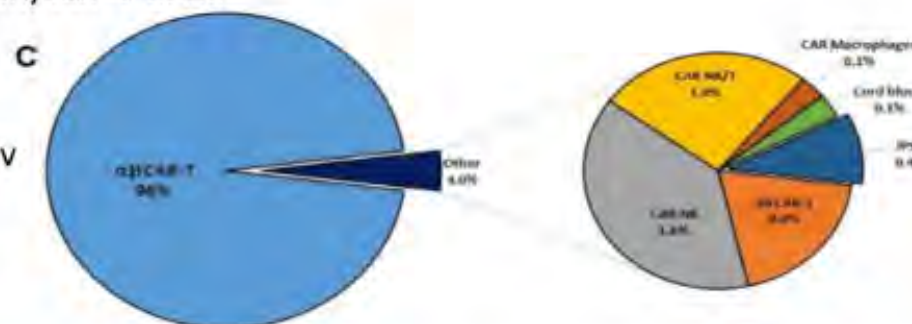
BBz, lenti

BBz, lenti

NCI- <https://www.cancer.gov/about-cancer/treatment/research/car-t-cells>



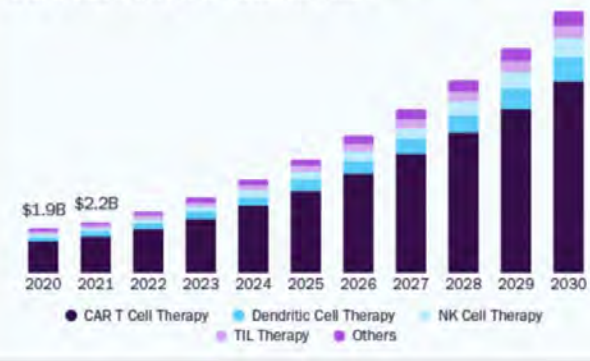
CD19 CAR trials account for 41% of 730 CART trials listed at clinicaltrials.gov (March 2022)  
*Globerson-Levin, Eur J Immunol, 2021*



MUCH OF THIS SUCCESS IS DUE TO CELLULAR IMMUNOTHERAPIES

**U.S. Cellular Immunotherapy Market**

size, by therapy type, 2020 - 2030 (USD Billion)



GRAND VIEW RESEARCH

**19.9%**  
U.S. Market CAGR,  
2022 - 2030

Source:  
www.grandviewresearch.com

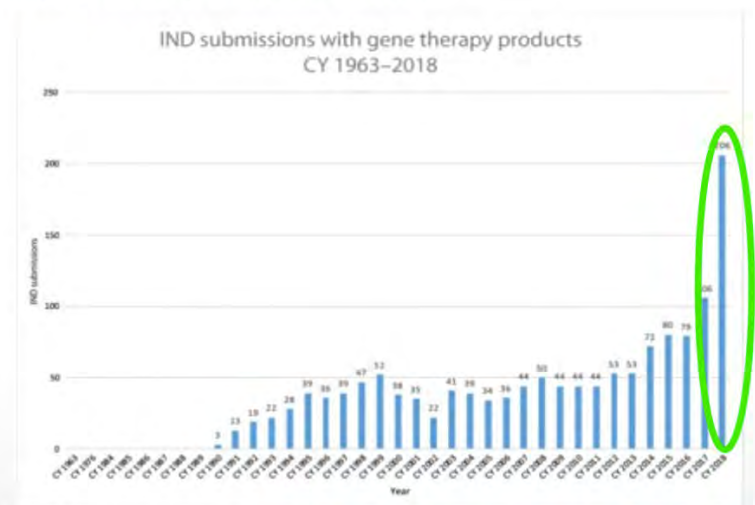
KEY DRIVERS OF MARKET GROWTH:

- Development of advanced cell-based therapies
  - 1,483 clinical trials in 2020
- Rising prevalence of cancer
  - 19.2M new cases in 2020, projected 24.5M by 2030
- Increasing R&D investment by key companies
- Increasing number of product approvals
- Growing adoption of cell-based immunotherapies



GENE & CELL THERAPY IS "BOOMING"

- In 2020, \$19.9 billion in financing



"By 2025, we predict that the FDA will be approving 10 to 20 cell and gene therapy products a year based on an assessment of the current pipeline and the clinical success rates of these products."<sup>4</sup> Dr. Scott Gottlieb

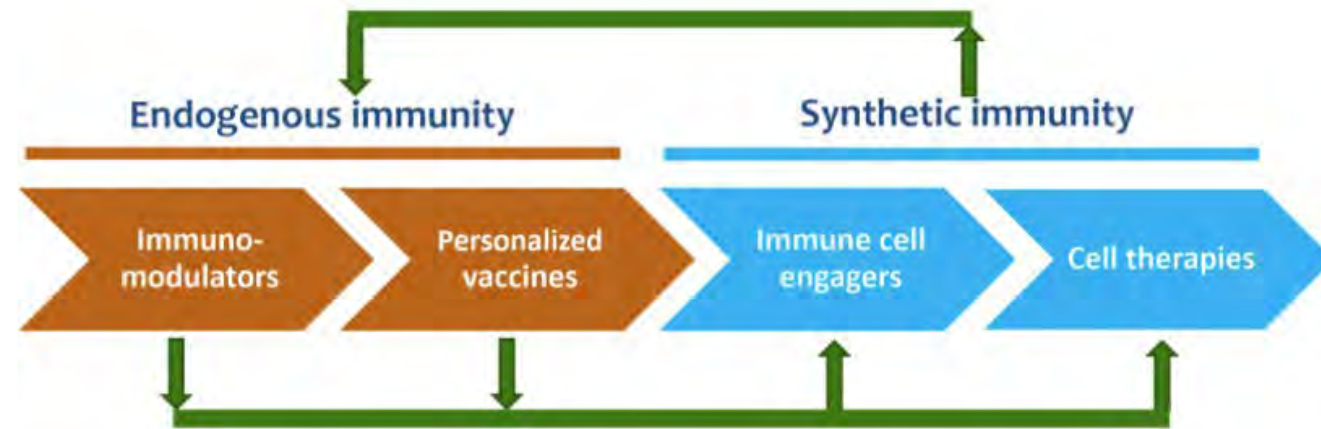
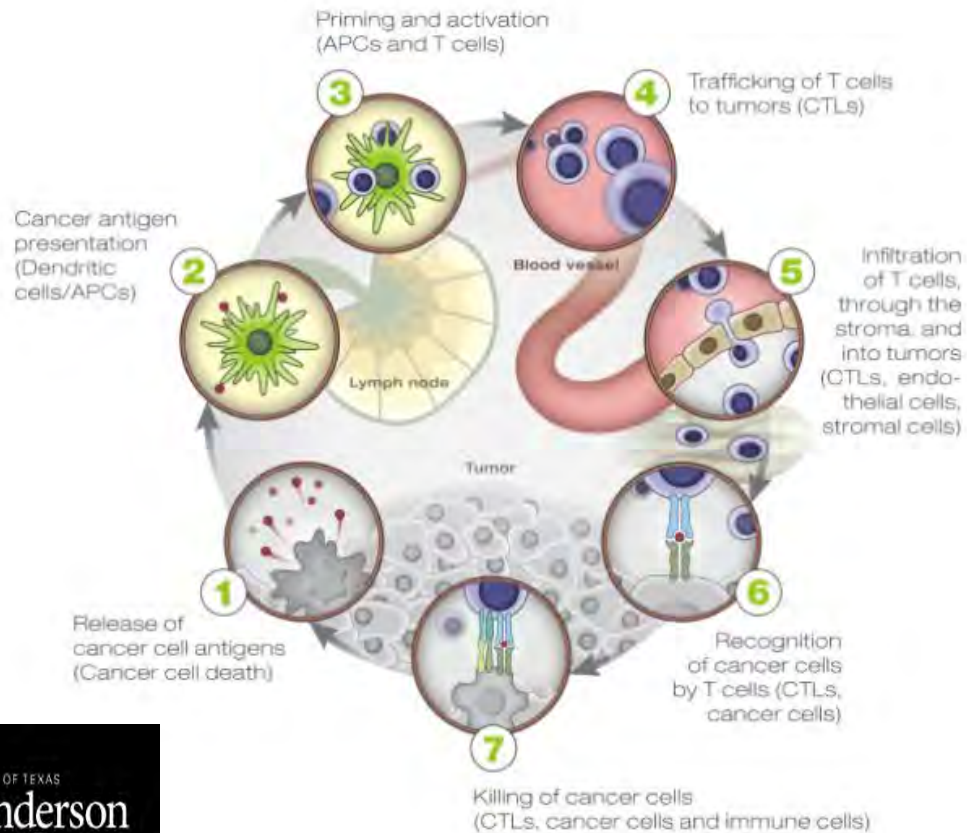
FDA Commissioner, 2017 - 2019





# The future: deploying multiple modalities to address both the intrinsic and extrinsic challenges to T cell\* function

\* or other effector cell

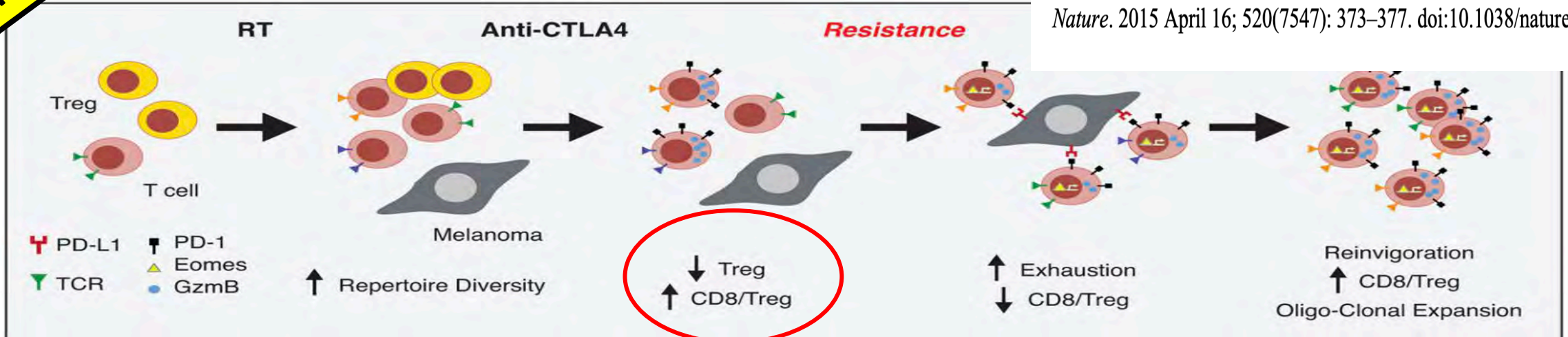


A 3D rendering of a puzzle. Most pieces are grey, but one piece in the center-right is bright red and stands out. The puzzle is set against a dark grey background that transitions to white on the right side.

How does Radiation fit  
in this complicated  
landscape?

Combining radiation and anti-CTLA4 or anti-PD1 immunotherapy for the treatment of melanoma.

Priming



Nature. 2015 April 16; 520(7547): 373–377. doi:10.1038/nature14292.

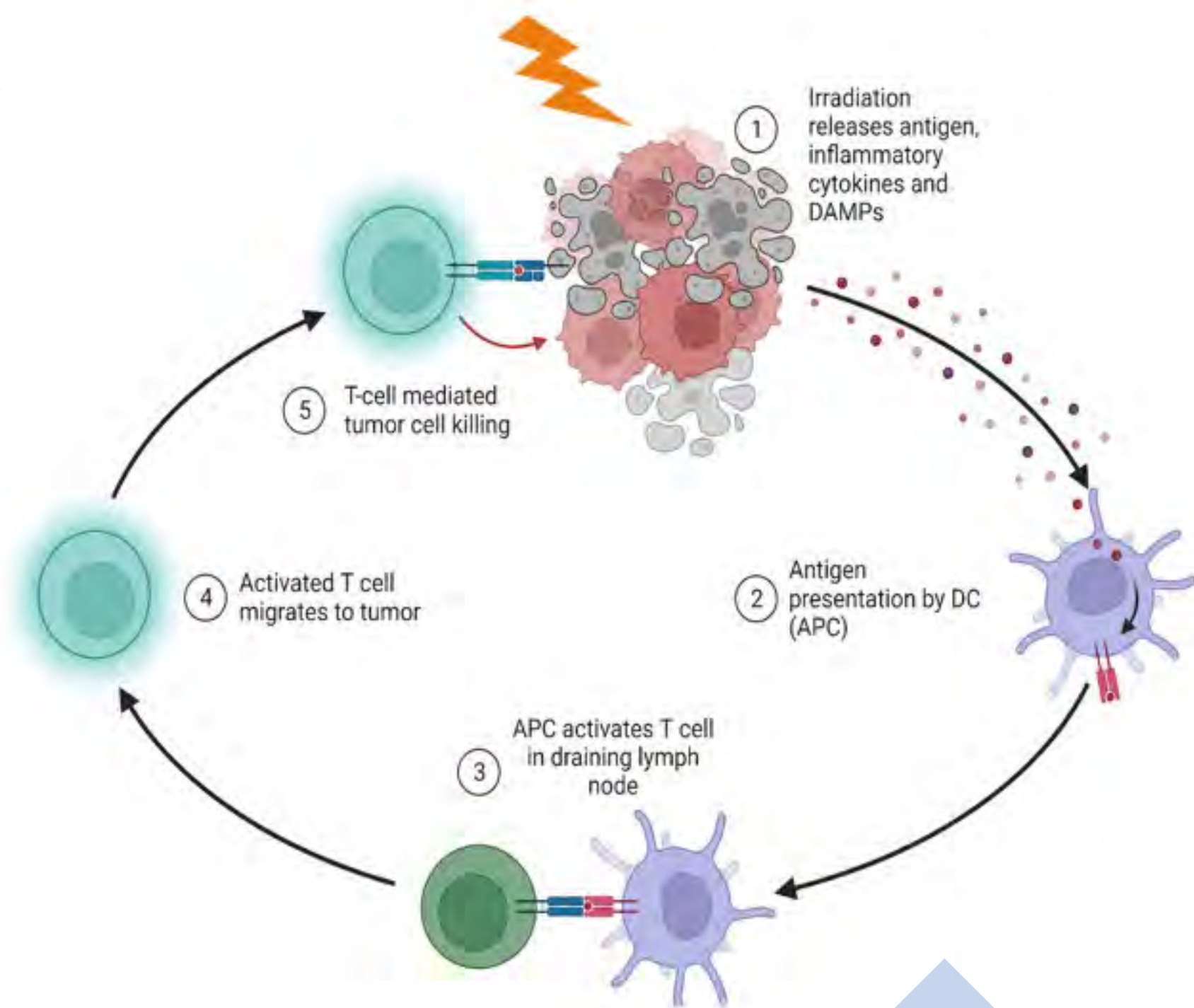
the University of Pennsylvania group demonstrated the importance of timing and sequence

After the initial response of combining anti-CTLA4 antibodies with radiation-----

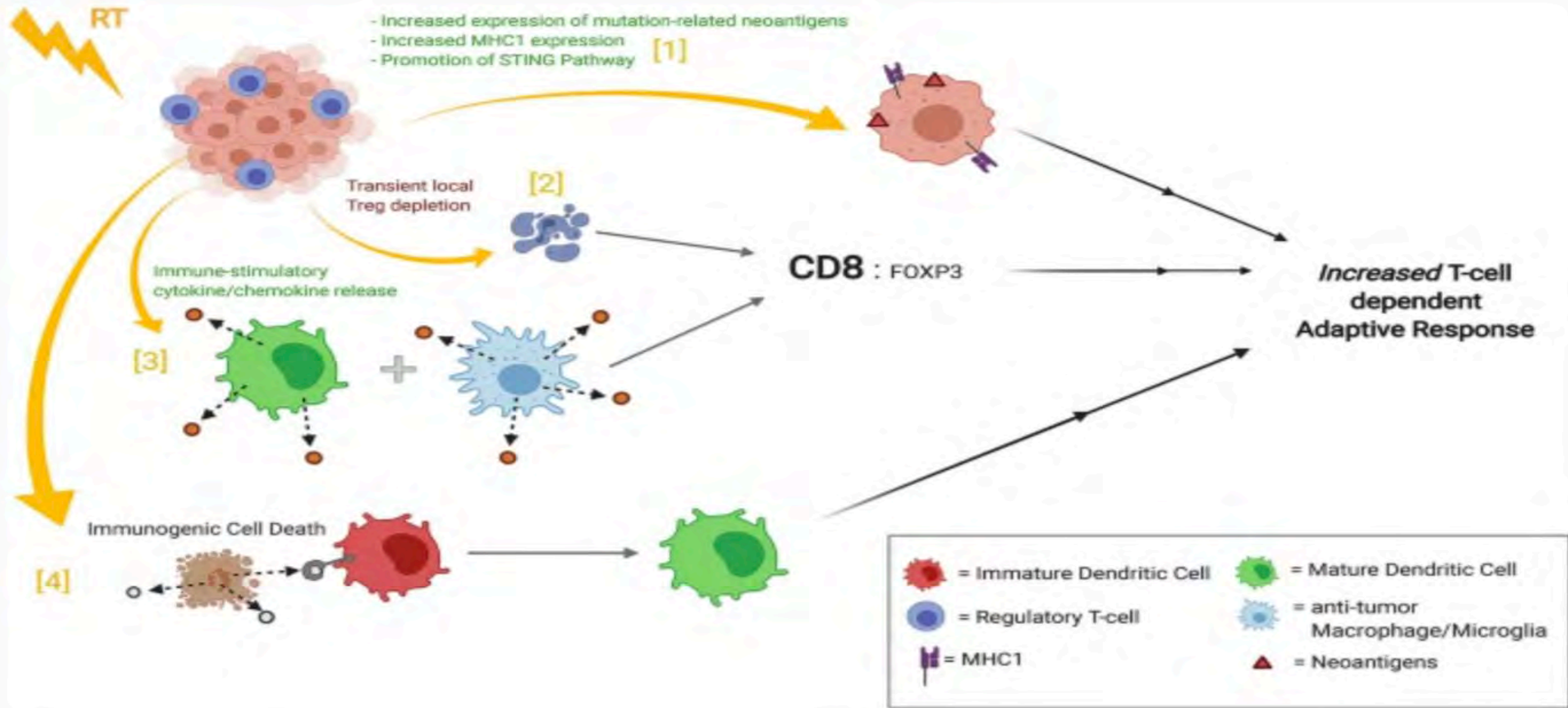
Resistance developed by upregulation of PDL1 leading to T-cell exhaustion\_\_\_\_\_

Resistance was reversed by adding PDL1 blockade.

# Priming



**Fig. 1**



**Strategies that improve antigen presentation:**

- Induce epitope spreading,
- Perpetuate existing antitumor T cell responses
- Combat tumor antigen escape.

What is the Available evidence?

# Radiotherapy and Immunogenic Cell Death

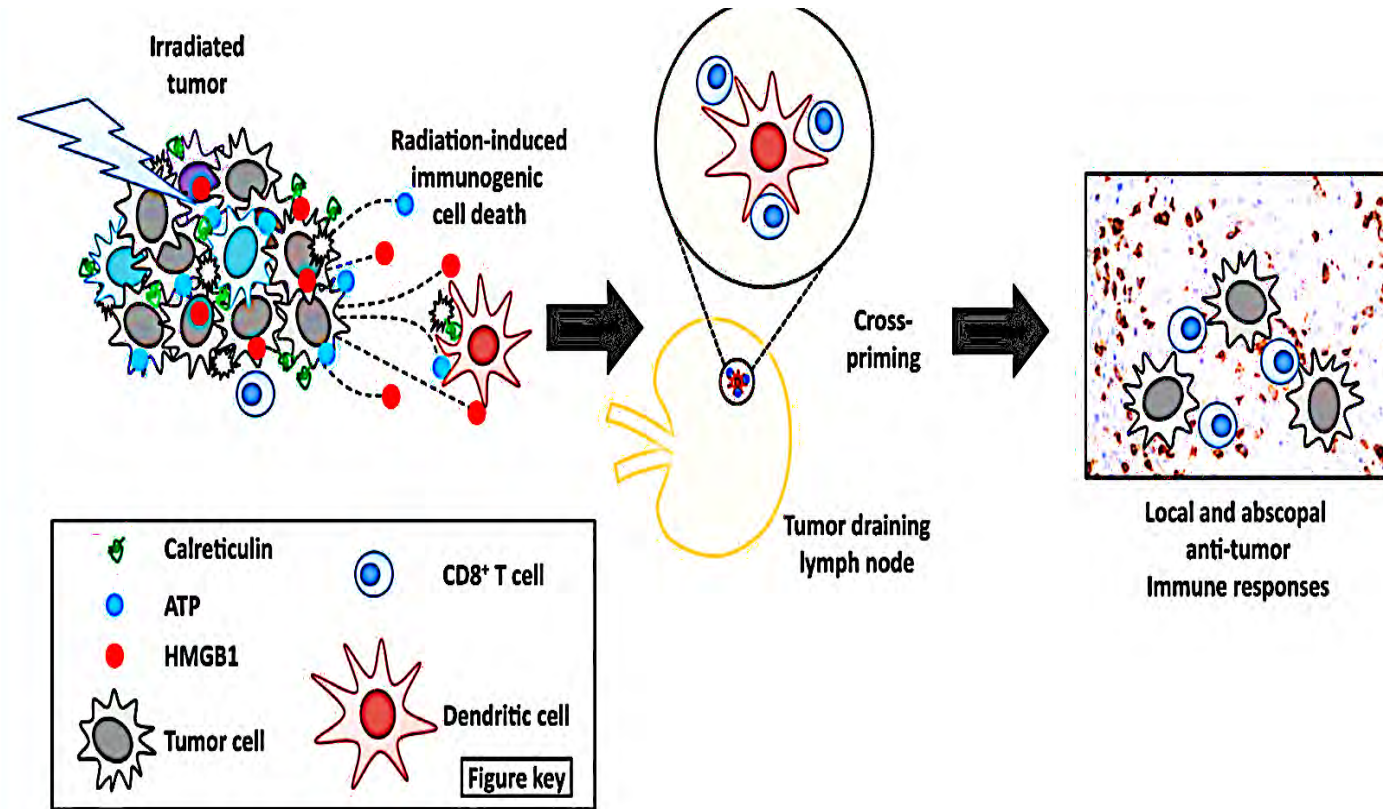


Seminars in  
**RADIATION  
ONCOLOGY**

Encouse B. Golden, MD, PhD,<sup>†,‡,§</sup> and Lionel Apetoh, PhD<sup>†,‡,§</sup>

## Immunogenic cell death= Priming

The dispersion of radiotherapy-induced immune-stimulating tumor antigens released from dying tumor cells into the surrounding milieu



T cell surface translocation DC the “eat me signal”

CD 47 DC (“do not eat me signal”) expressed by tumors

# Supportive evidence

# What is the Priming Dose of Radiation?

## Low-Dose Irradiation Programs Macrophage Differentiation to an iNOS<sup>+</sup>/M1 Phenotype that Orchestrates Effective T Cell Immunotherapy

Felix Klug,<sup>1,11</sup> Hridayesh Prakash,<sup>1,2,4,11</sup> Peter E. Huber,<sup>3,11,\*</sup> Tobias Seibel,<sup>1,11</sup> Noemi Bender,<sup>1</sup> Niels Halama,<sup>4</sup> Christina Pfirschke,<sup>1</sup> Ralf Holger Voss,<sup>1</sup> Carmen Timke,<sup>3</sup> Ludmila Umansky,<sup>1</sup> Kay Klapproth,<sup>5</sup> Knut Schäkel,<sup>2</sup> Natalio Garbi,<sup>6,10</sup> Dirk Jäger,<sup>8</sup> Jürgen Weitz,<sup>3</sup> Hubertus Schmitz-Winnenthal,<sup>3</sup> Günter J. Hämmerling,<sup>1</sup> and Philipp Bechhoeve<sup>1,\*</sup>



- normalization of aberrant vasculature
- recruitment of tumor-specific T cells in human pancreatic carcinomas
- **T-cell-mediated tumor rejection**  
prolonged survival in otherwise immune refractory spontaneous and xenotransplant mouse tumor models.

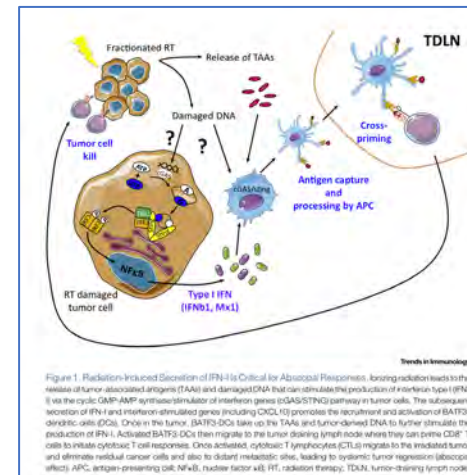
Trends in Immunology

CellPress REVIEWS

Review

## Immunological Mechanisms Responsible for Radiation-Induced Abscopal Effect

Maria E. Rodríguez-Ruiz,<sup>1,2,5,6</sup> Claire Vanpouille-Box,<sup>2,5</sup> Ignacio Melero,<sup>1</sup> Silvia Chiara Formenti,<sup>2,3</sup> and Sandra Demaria<sup>2,3,4,\*</sup>



### Ionizing Radiation has multiple immune-modulatory effects:

- Production of IFN-β, DCs infiltrating the tumor
- **promoting the cross-presentation of tumor antigens to CD8 T cells**

### T cell infiltration following radiation

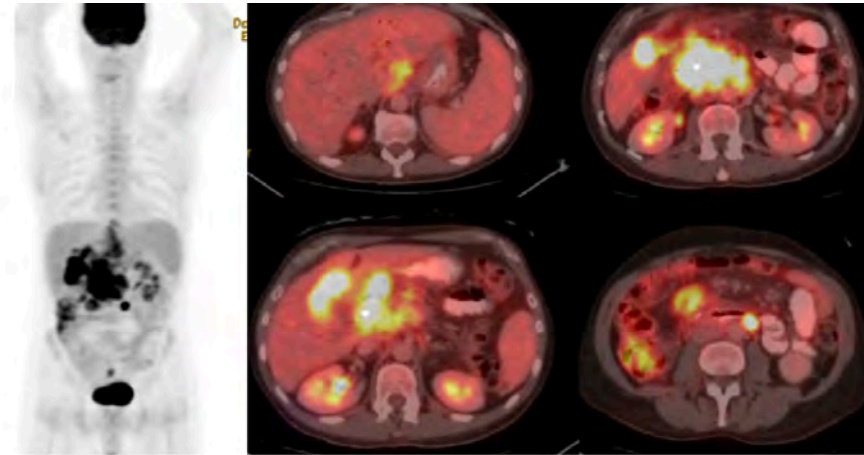
- IFN-stimulated genes that encode the chemokines
  - CCL5, CXCL16 (C-X-C motif chemokine ligand 16), CXCL10



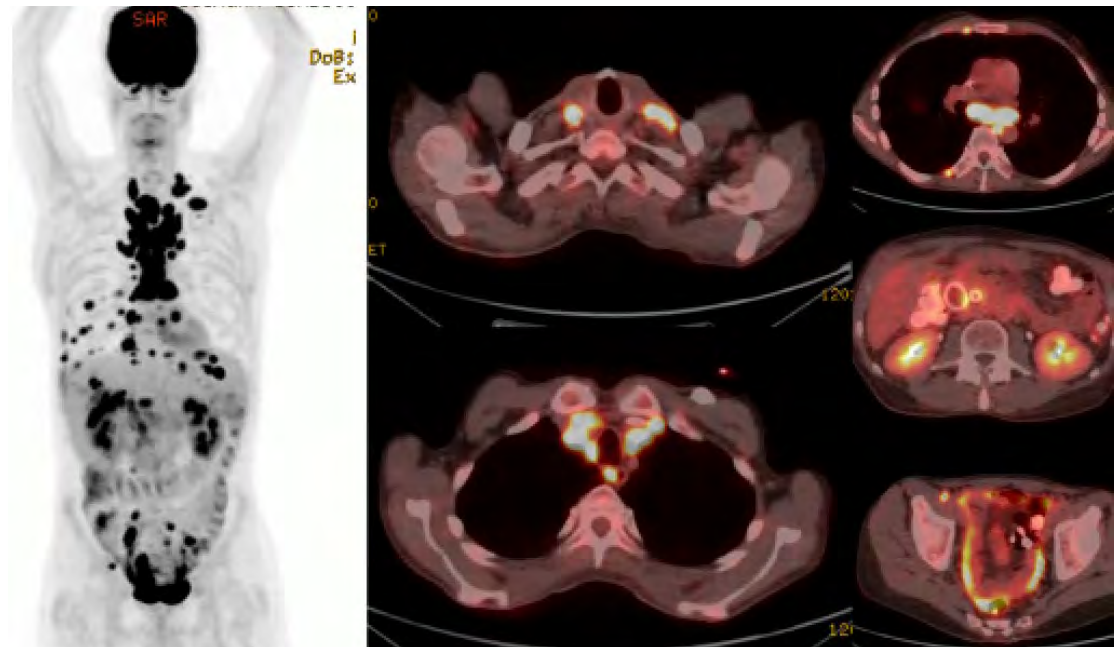
How does this translate clinically?

Are we sure that the immunological changes equate a clinical benefit ?

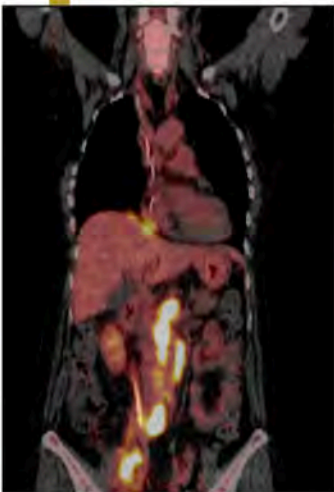
An example of what we see in clinic  
Confirming that radiation does stimulate the immune system



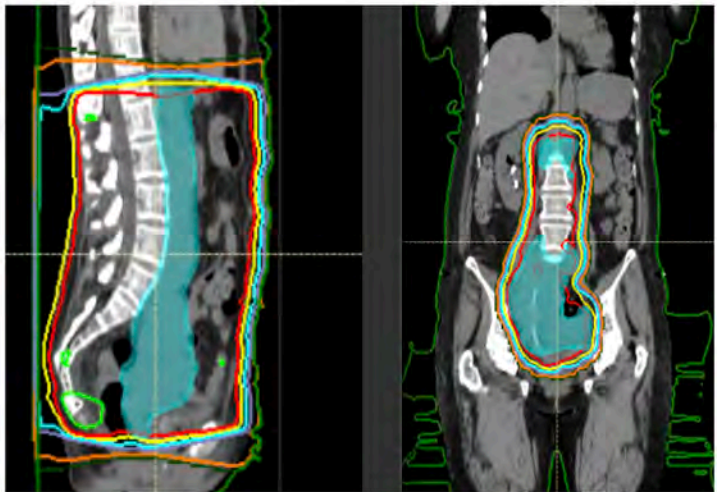
Before Radiation



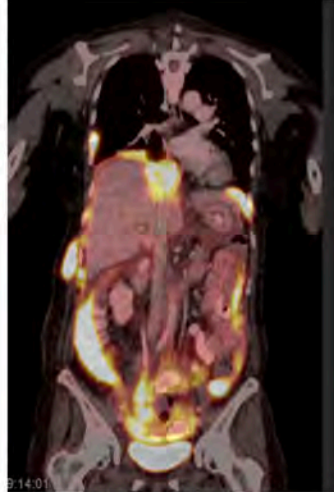
**Day 7 form start of RT: flare of nodes outside of RT**



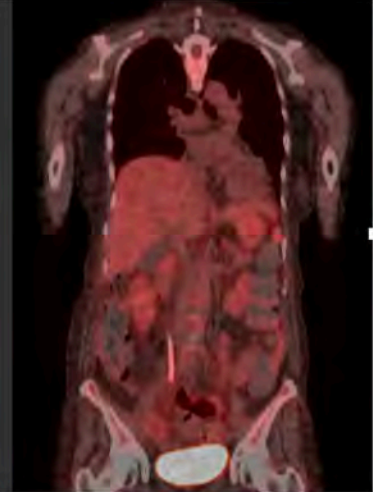
Presentation to RO mass along the PA area to pelvis



Field of radiation treating all but the diaphragmatic lesion



PET done before proceeding to CAR T Infusion suspected relapse everywhere except in the Irradiated field



Proceeded to Car T and achieved a durable complete remission

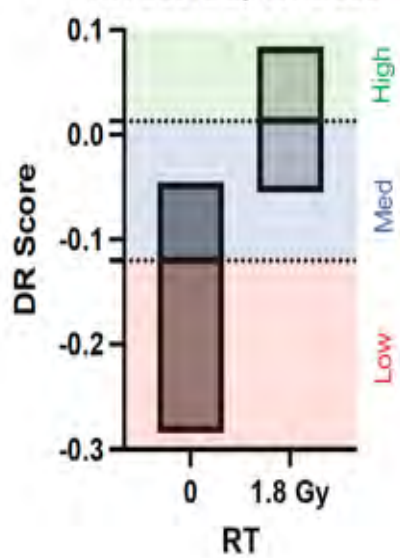
# Exploring low-dose Radiation as low as 1.8 Gy

RESEARCH ARTICLE | APRIL 24, 2023

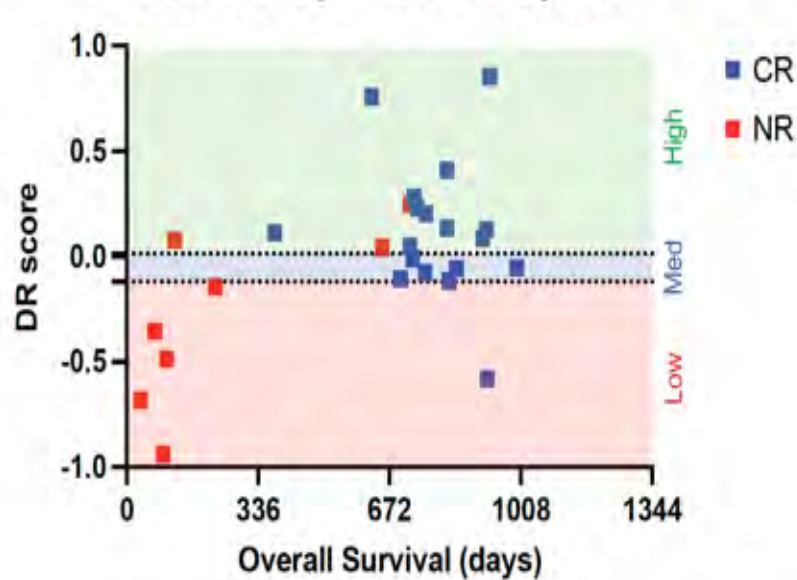
## Intrinsic tumor resistance to CAR T cells is a dynamic transcriptional state that is exploitable with low-dose radiation

Alexander B. Kim, Ssu-Yu Chou, Solomon Kang, Eric Kwon, Matthew Inkman, Jeffrey Szymanski, Neal Andruska, Cian Colgan, Jin Zhang, Joanna C Yang, Nathan Singh, Carl DeSelm

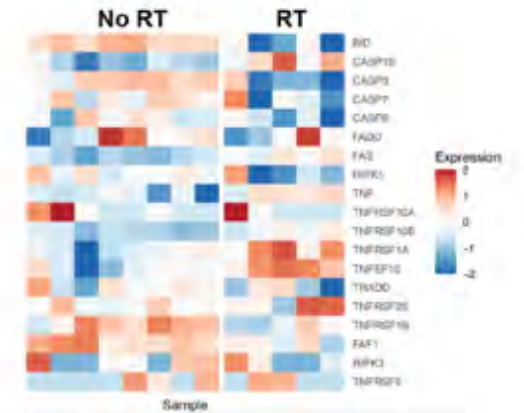
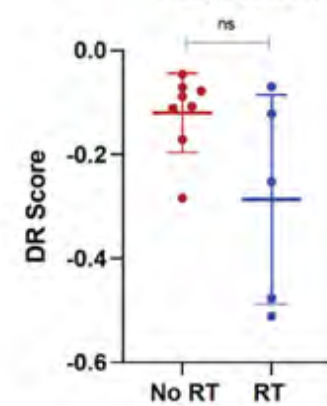
a DR Score by RT Dose



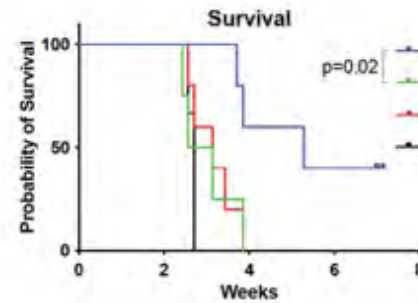
b DR score by time and response



7 days post RT

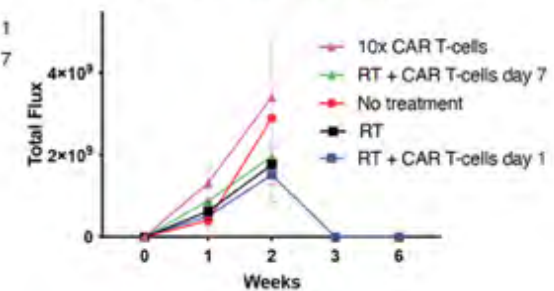


c



d

RT Timing Tumor Burden



- LD-TBI plus CAR T cells on day 0 exhibited superior tumor control and OS (median: 110 days) as compared to all other groups.
- considerable expansion of circulating T cells

ARTICLE OPEN

Check for updates

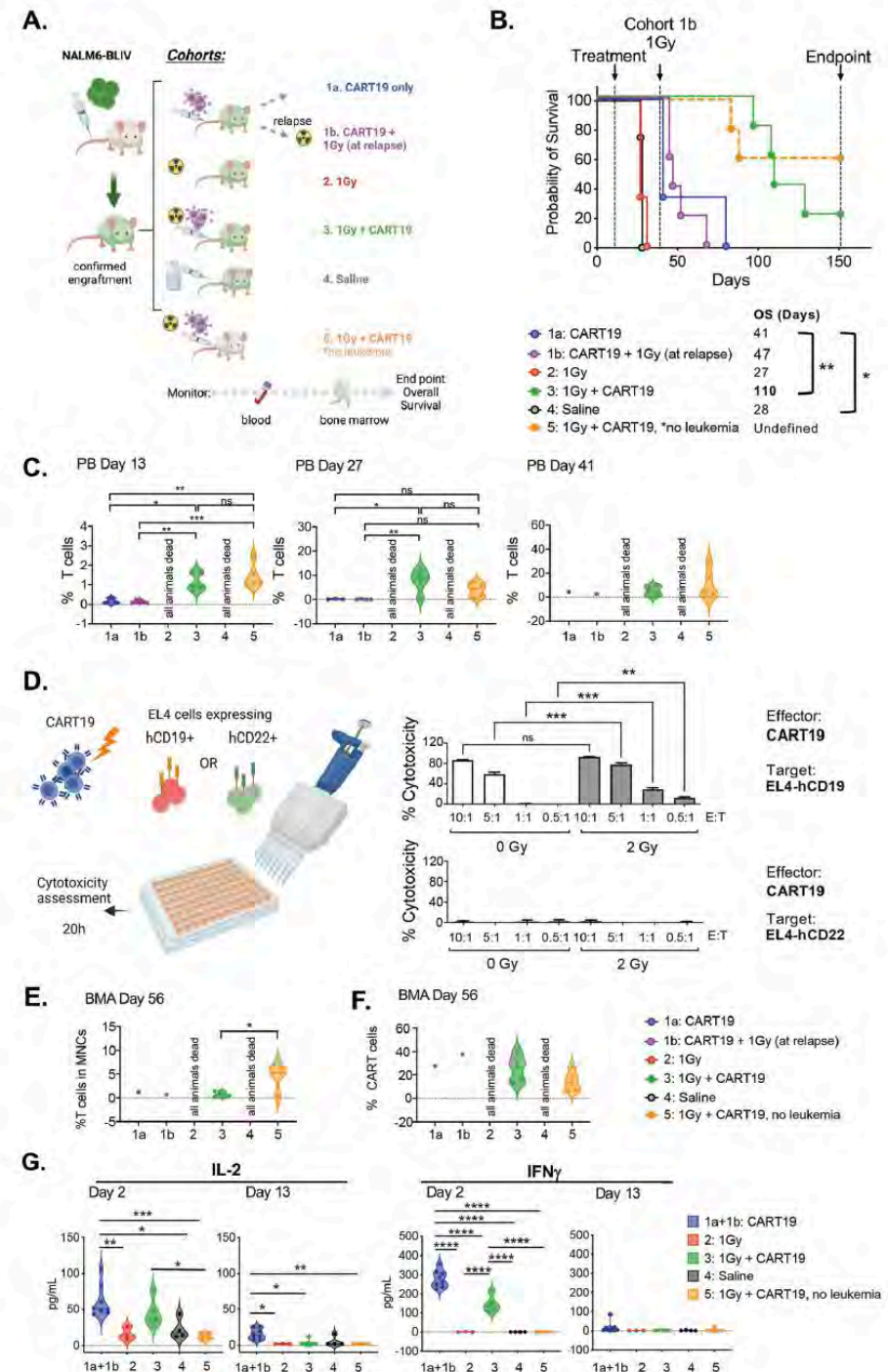
# Radiation therapy improves CAR T cell activity in acute lymphoblastic leukemia

Mayumi Sugita<sup>1,8</sup>, Takahiro Yamazaki<sup>2,8</sup>, Mohammad Alhomoud<sup>1</sup>, Jérémie Martinet<sup>1,3,4</sup>, Jean-Baptiste Latouche<sup>3,4</sup>, Encouse Golden<sup>2</sup>, Olivier Boyer<sup>3,4</sup>, Koen Van Besien<sup>1</sup>, Silvia C. Formenti<sup>2,5</sup>, Lorenzo Galluzzi<sup>2,5,6,9</sup> and Monica L. Guzman<sup>1,5,6,7,9</sup>

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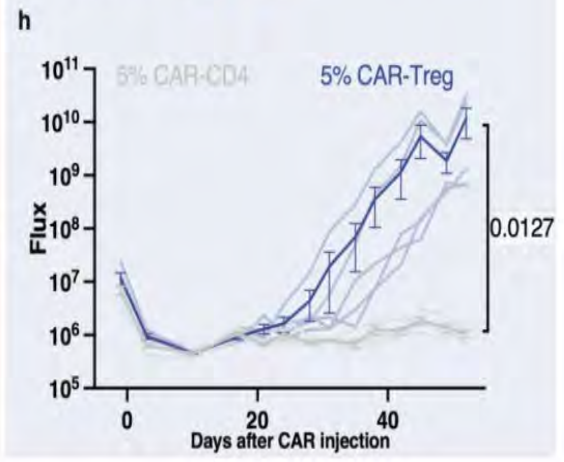
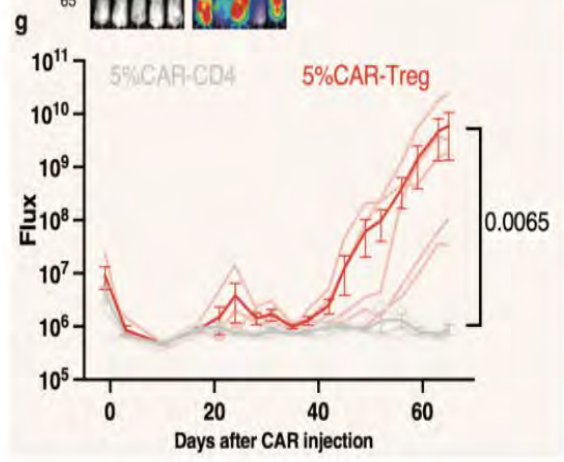
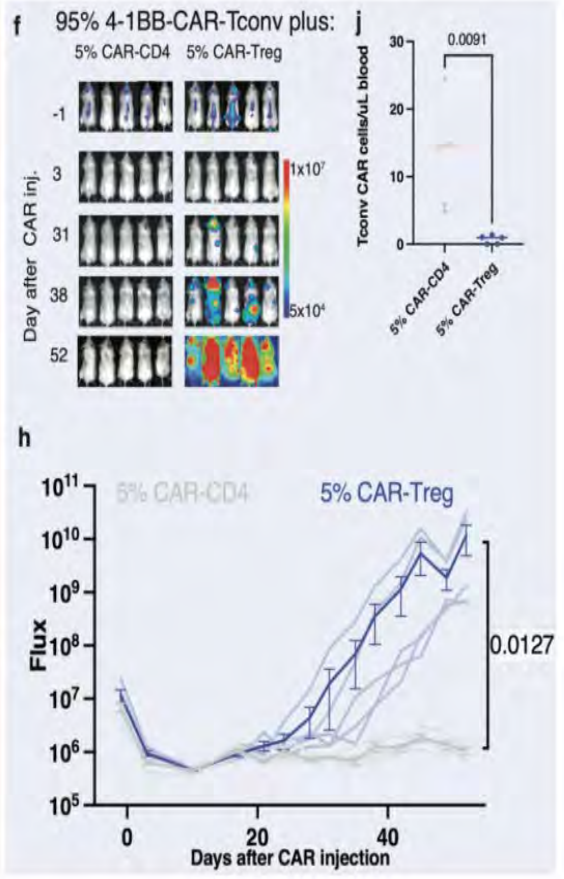
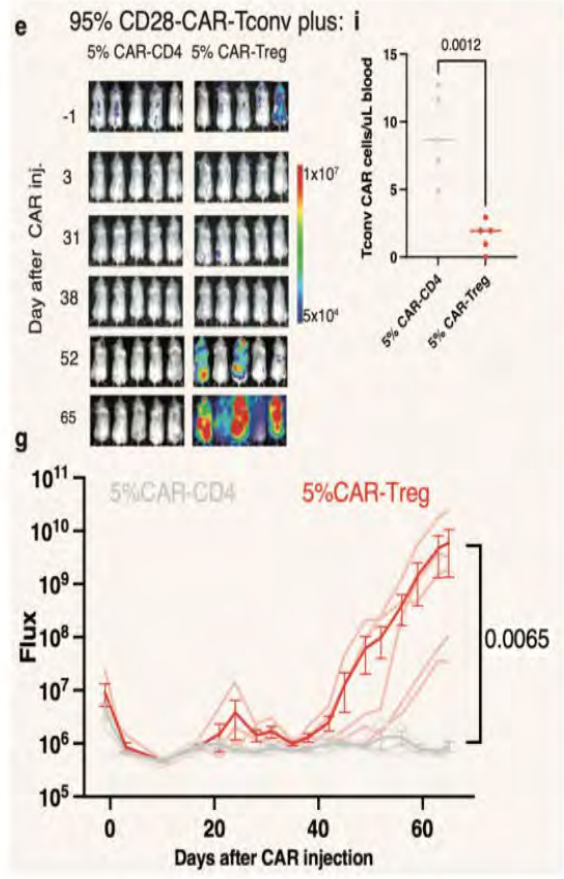
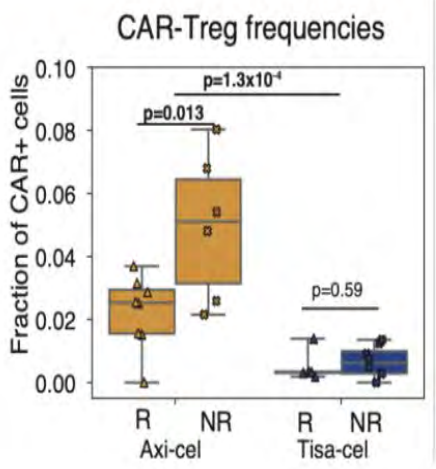
Low-dose total body irradiation (LD-TBI) superior expansion of CAR T cells in vivo.

RT to elicit death receptor (DR) expression by malignant cells enabling some degree of CAR-independent tumor killing



**Priming**

# CAR-Tregs may have a role in depressing responses



Elevations in CAR-T regulatory cells among non-responders to Axi-cel

- capable of suppressing conventional CAR-T cell expansion
- driving late relapses in an in vivo model.

The capacity for even small increases in CAR-T regulatory cells to drive relapse.

# Ionizing radiation modulates the phenotype and function of human CD4+ induced regulatory T cells

Samantha S. Beauford, Anita Kumari and Charlie Garnett-Benson\*



*Created an Opportunity  
for Radiation*

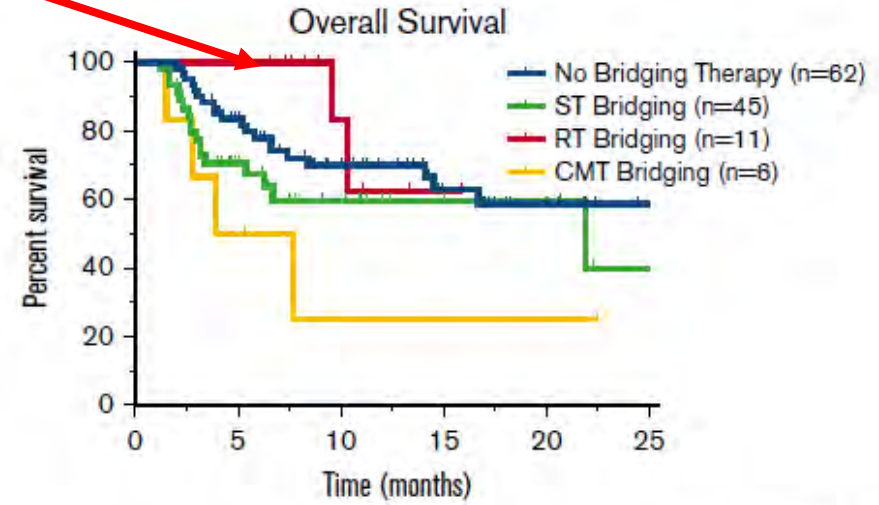
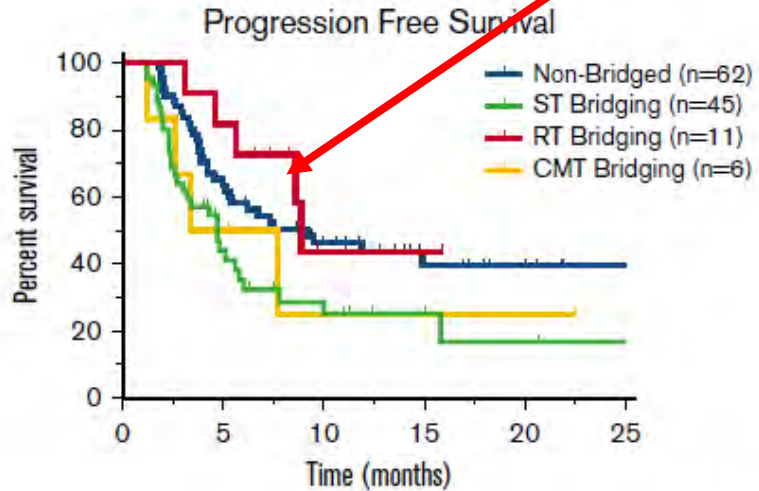
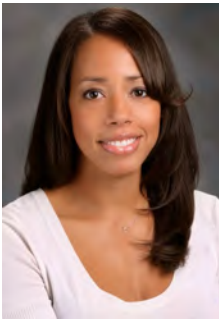
**Conclusions:** Our findings demonstrate that while human T<sub>REG</sub> cells are more resistant to radiation-induced death, treatment causes downregulation of Foxp3 expression, as well as modulation in the expression of T<sub>REG</sub> signature molecules associated with suppressive activity. Functionally, irradiated TGF- $\beta$ 1-induced T<sub>REGS</sub> were less effective at inhibiting CD8+ T cell proliferation. These data suggest that doses of radiotherapy in the hypofractionated range could be utilized to effectively target and reduce T<sub>REG</sub> activity, particularly when used in combination with cancer immunotherapies.



Does the preclinical data translate into a clinical benefit?



# Radiation priming is effective



No Bridging	62	37	22	11	5	1
ST Bridging	45	16	8	4	3	1
RT Bridging	11	10	3	2	0	0
CMT Bridging	6	4	2	2	1	0

**No bridging vs ST  $p=0.01$**   
 No bridging vs RT  $p=0.52$   
 No bridging vs CMT  $p=0.36$   
**ST vs RT  $p=0.05$**   
 ST vs CMT  $p=0.78$   
 RT vs CMT  $p=0.17$

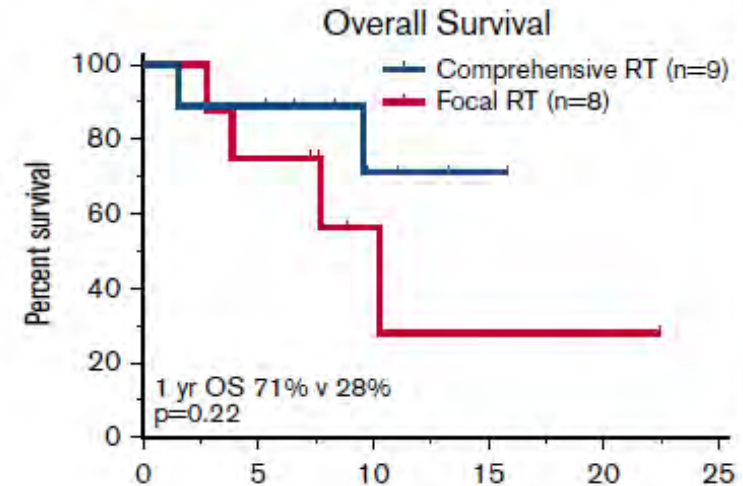
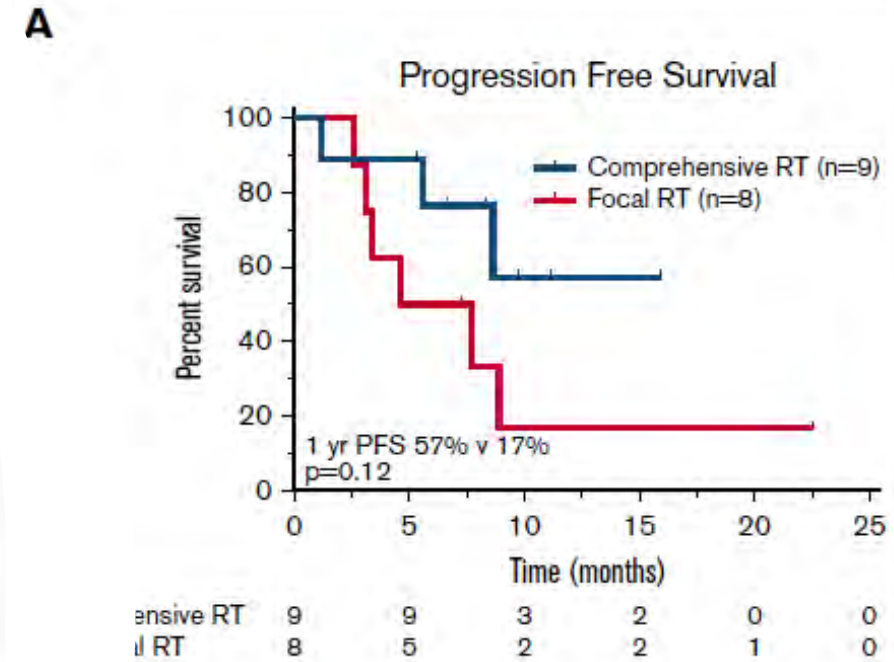
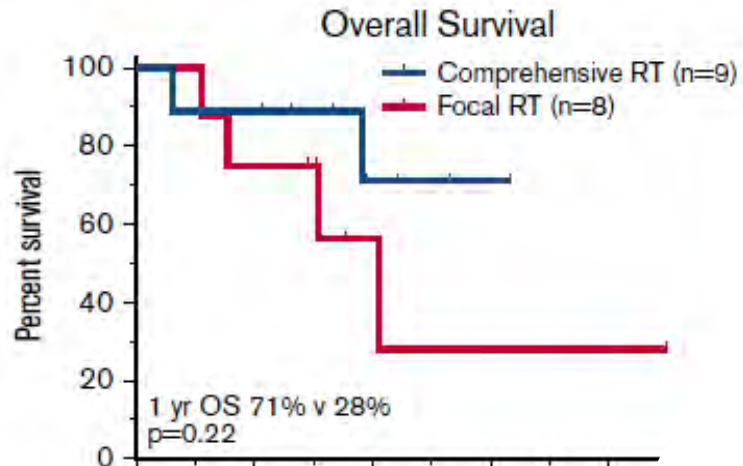
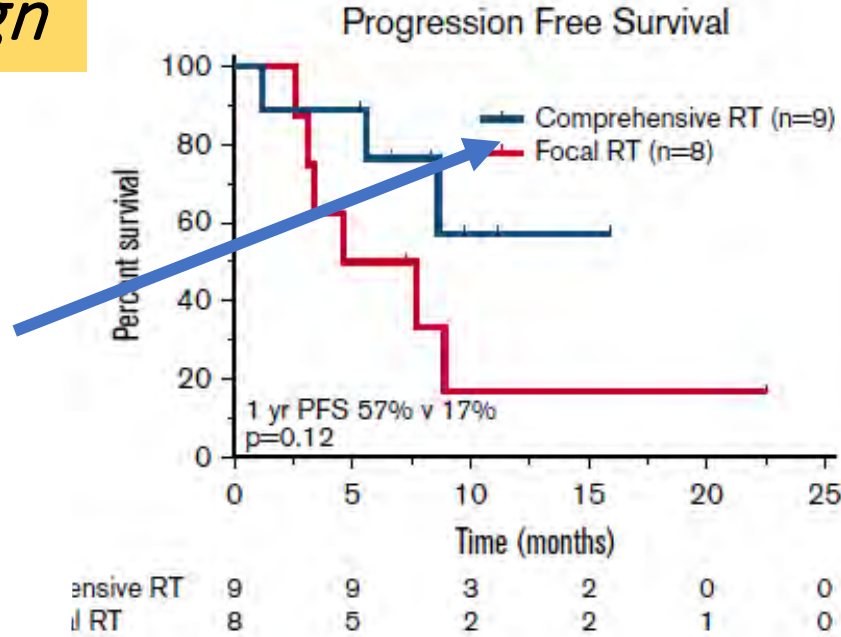
No Bridging	62	48	30	16	7	2
ST Bridging	45	26	12	5	5	2
RT Bridging	11	11	5	2	1	0
CMT Bridging	6	4	2	2	2	0

No bridging vs ST  $p=0.17$   
 No bridging vs RT  $p=0.40$   
**No bridging vs CMT  $p=0.03$**   
 ST vs RT  $p=0.15$   
 ST vs CMT  $p=0.30$   
**RT vs CMT  $p=0.02$**

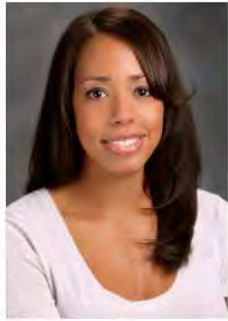
- Compared to patients that did not receive BT
  - 1-year PFS 44% for no BT vs 25% for ST cohort ( $p=0.01$ )
- Patients treated with CMT had a poor outcome
  - 1-year OS 25% and median OS of 3.9 months

# Impact of RT Field Design

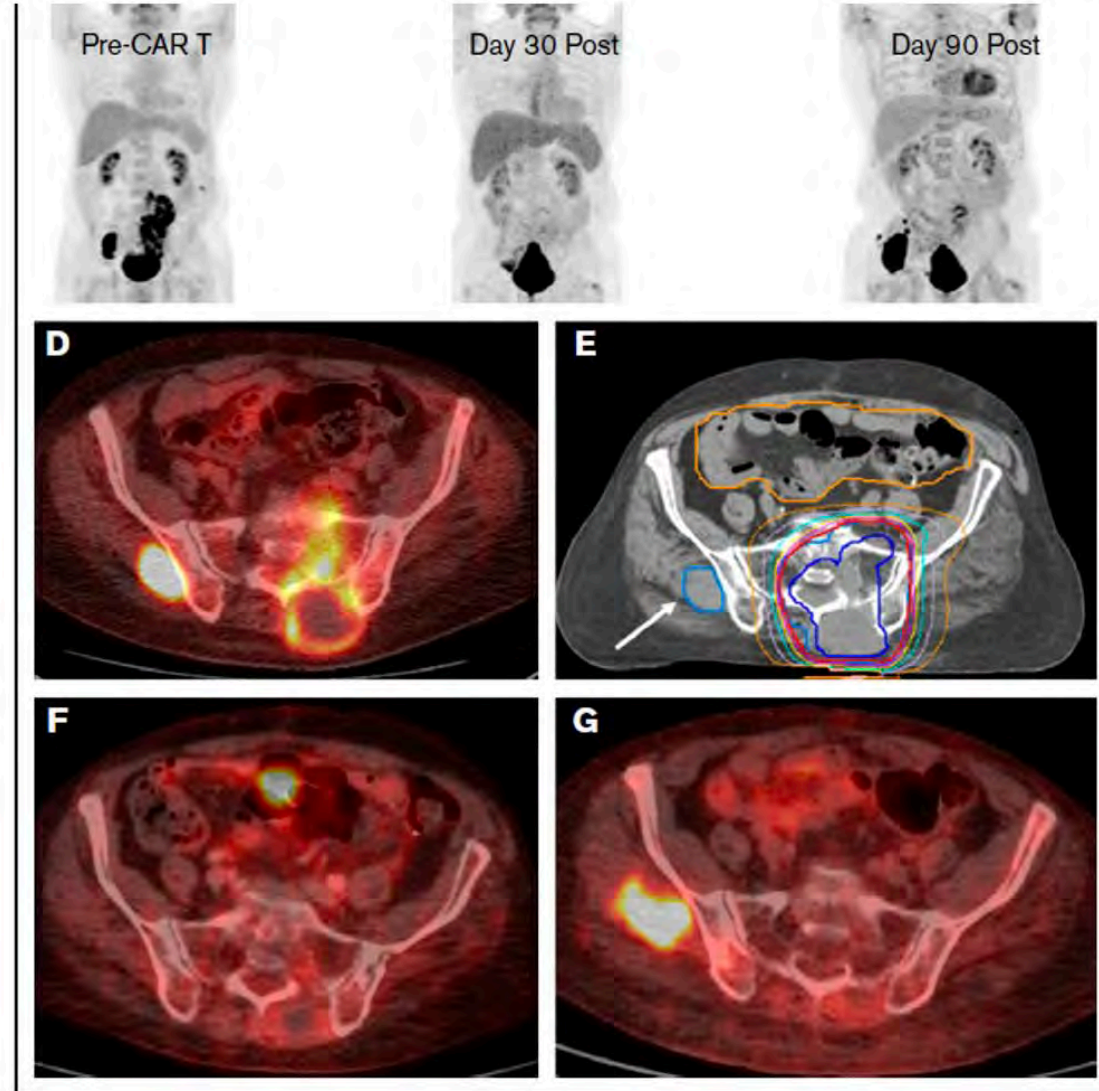
- Nine patients comprehensive RT field (CMT, n=2; RT, n=7)
- 8 patients received focal RT fields (CMT, n=4; RT, n=4)
  - 6 progressed or relapsed
  - 3 at sites not included in the RT field



# Importance of RT Field Size for Bridging Therapy?

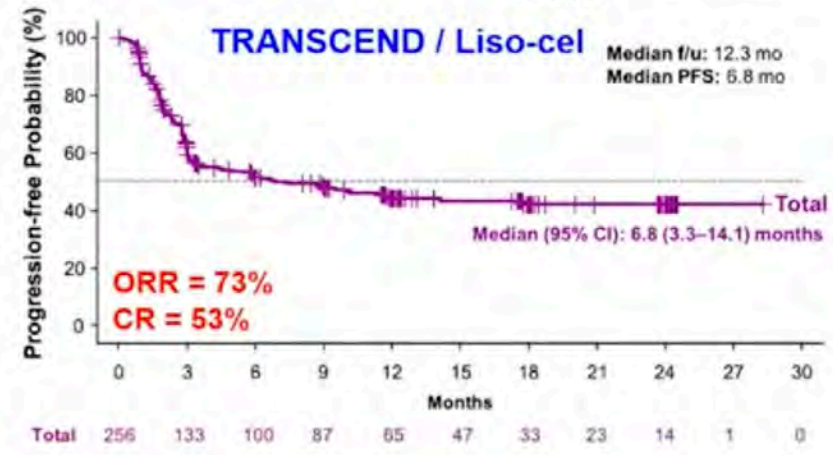
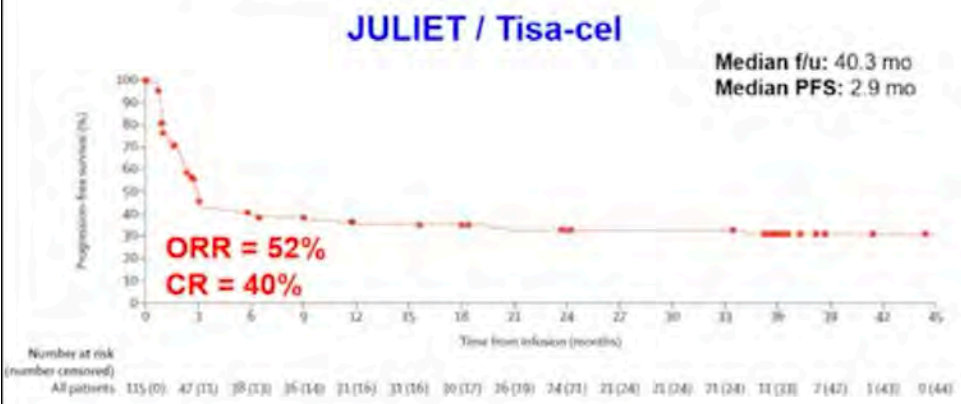
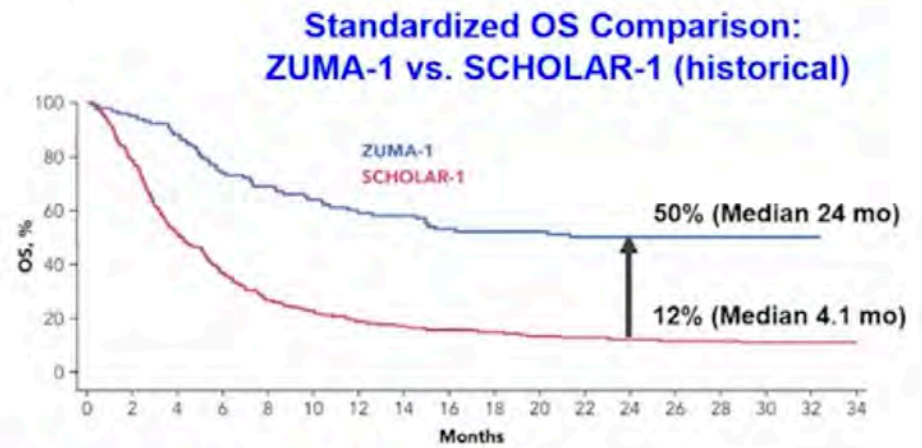
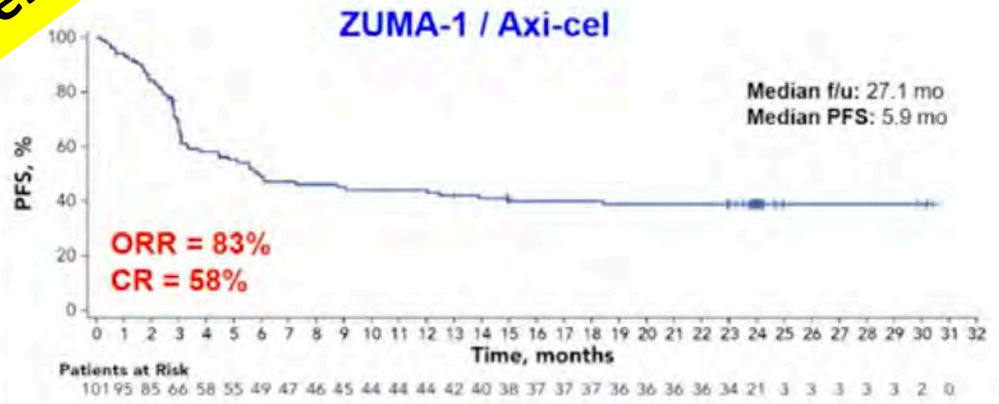


- Stage IV primary refractory HGBL-DH (MYC/BCL2) after DA R-EPOCH
- R-DHAP → progression
- P/w neurologic symptoms
- CAR-T with RT bridging
- Relapsed in un-irradiated site



**Avoiding further Chemotherapy**

# CD19 CART in $\geq 3^{\text{rd}}$ line LBCL



Neelapu et al. *N Eng J Med* 2017; Locke et al. *Lancet Oncol* 2019  
Schuster et al. *N Eng J Med* 2019; Schuster et al. *Lancet Oncol* 2021  
Abramson et al. *Lancet* 2020; Neelapu et al. *Blood Adv* 2021

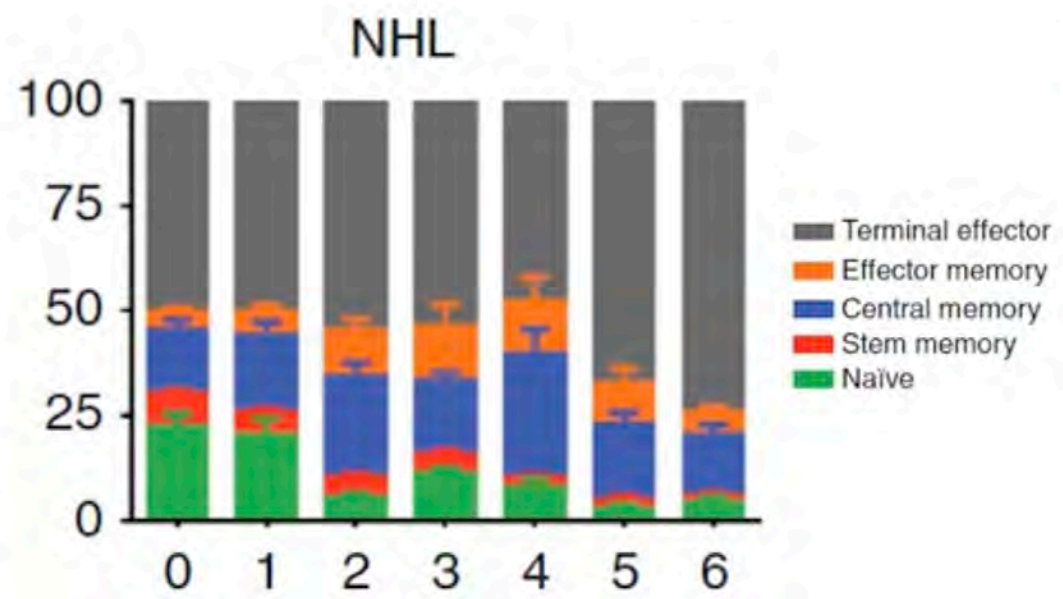
1. Better than standard of care
2. Those who get CR it is durable

Adopted from Dr. Sattva Neelapu

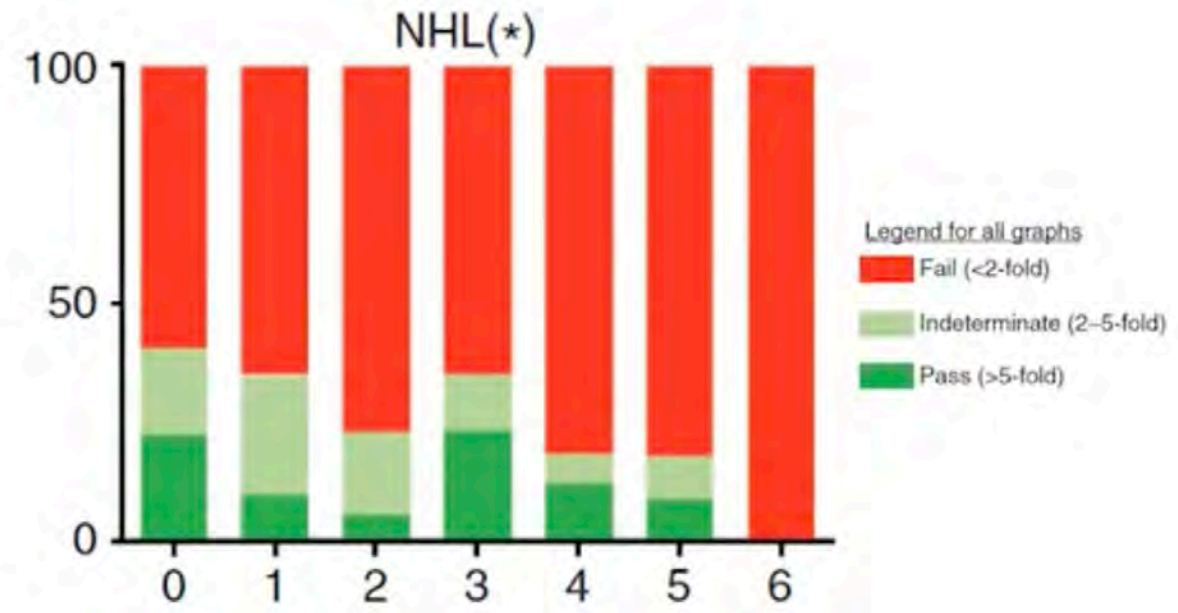
Avoiding Chemotherapy

# Chemotherapy impairs immune cell phenotype and function

## T-cell phenotype



## T-cell proliferation



Naïve and terminal effector are impaired by chemotherapy

Das et al. *Cancer Discov* 2019; 9(4): 492-499

Adopted from Dr. Sattva Neelapu

**Avoiding Chemotherapy**

# ZUMA 7 moving CAR T to second line

The NEW ENGLAND JOURNAL of MEDICINE

ORIGINAL ARTICLE

## Survival with Axicabtagene Ciloleucel in Large B-Cell Lymphoma

J.R. Westin, O.O. Oluwole, M.J. Kersten, D.B. Miklos, M.-A. Perales, A. Ghobadi, A.P. Rapoport, A. Sureda, C.A. Jacobson, U. Farooq, T. van Meerten, M. Ulrickson, M. Elsayy, L.A. Leslie, S. Chaganti, M. Dickinson, K. Dorritie, P.M. Reagan, J. McGuirk, K.W. Song, P.A. Riedell, M.C. Minnema, Y. Yang, S. Vardhanabhuti, S. Filosto, P. Cheng, S.A. Shahani, M. Schupp, C. To, and F.L. Locke, for the ZUMA-7 Investigators and Kite Members\*

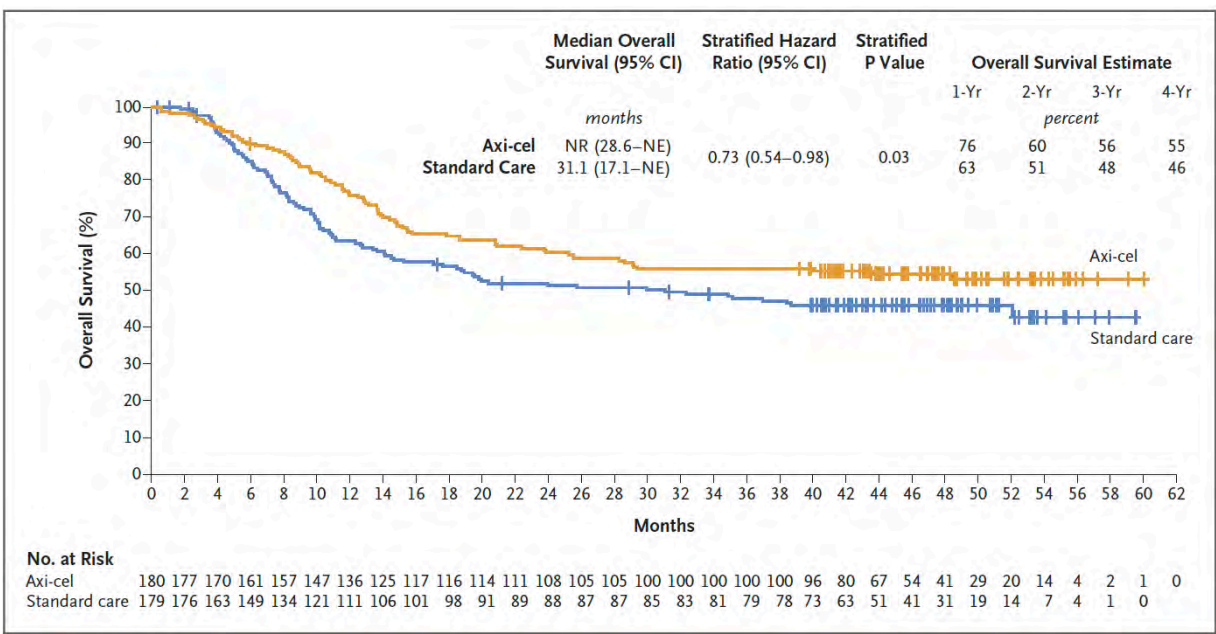


Figure 1. Overall Survival.

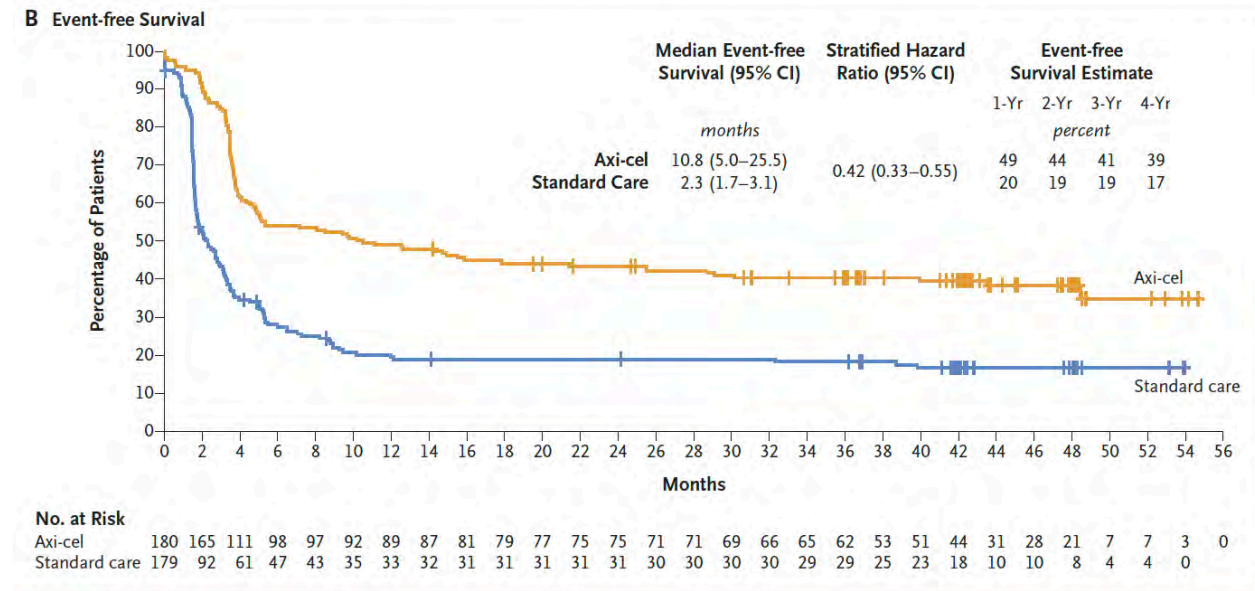
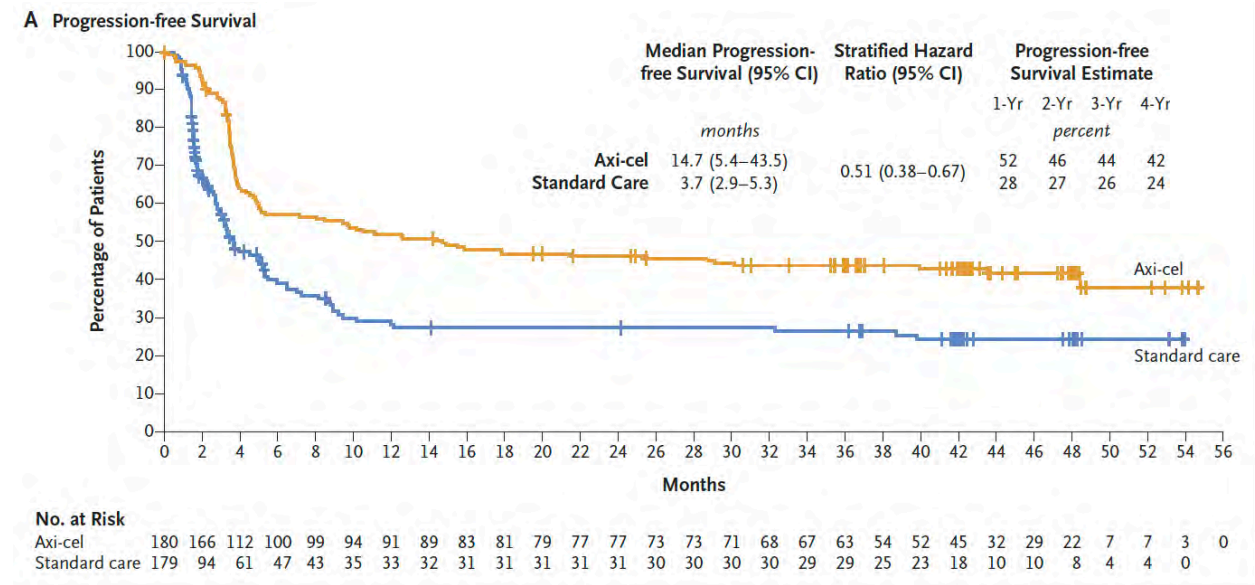


Figure 2. Progression-free Survival and Event-free Survival, as Assessed by the Investigator.

# Primary Mediastinal Lymphoma

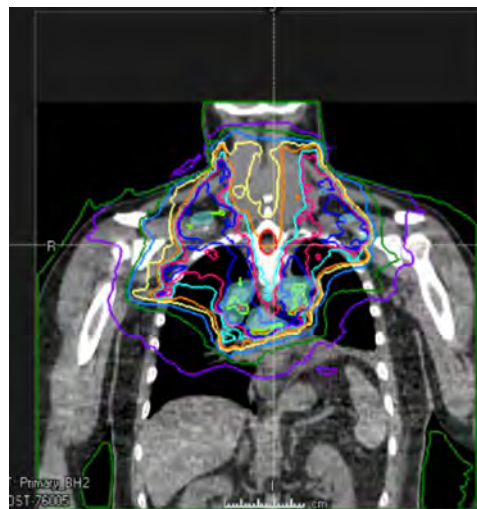
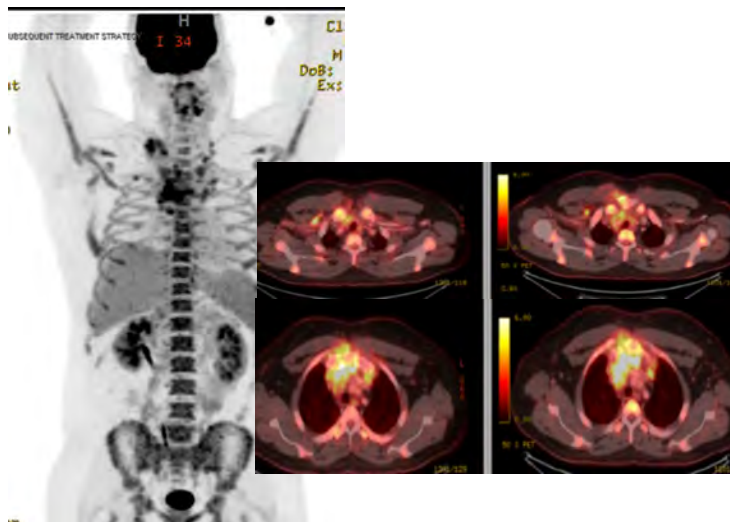
Example of heavily pretreated



BV-Nivo on 2020-0686 PACIFIC

Progression

DA-REPOCH x 4



Relapse ----- Hypercytotoxin

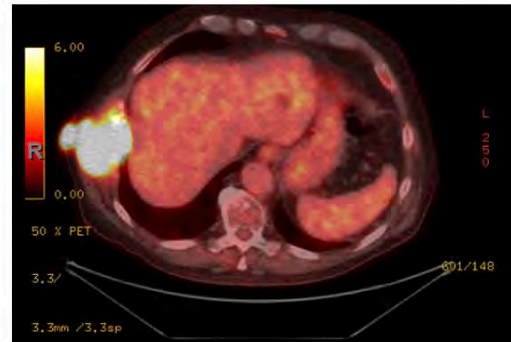
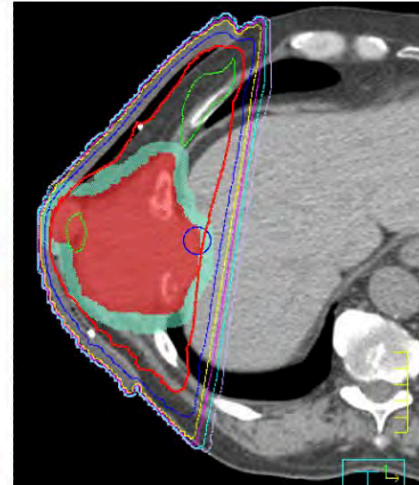
Radiation to 30 Gy

Post CAR T 10 months in CR

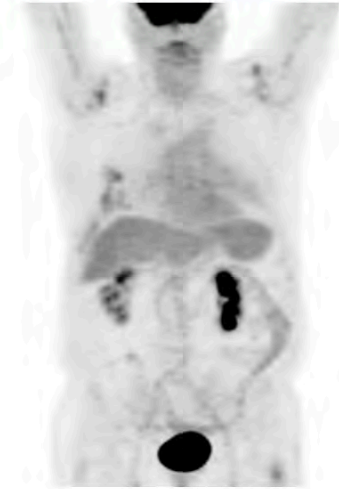
**Radiation a better  
Alternative to further  
chemotherapy**

Refractory after 6 RCHOP  
R-DHAP x 1 → initial  
response but rapid  
progression

Pre-RT/Pre-CART



Post 30-day PET-CT: CR  
with 5PS of 3

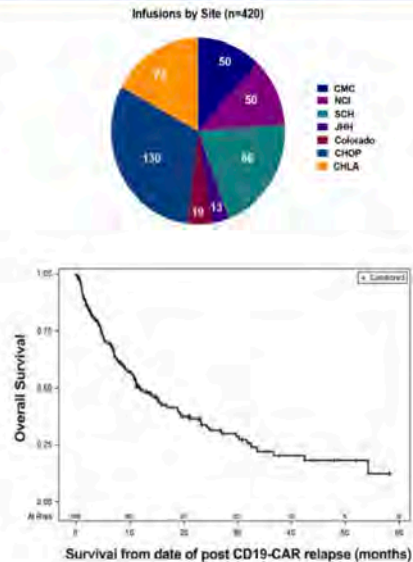




## Predicting relapse post CAR T An opportunity to improve the outcome

### Outcomes for post-CD19 CAR Relapse are Poor

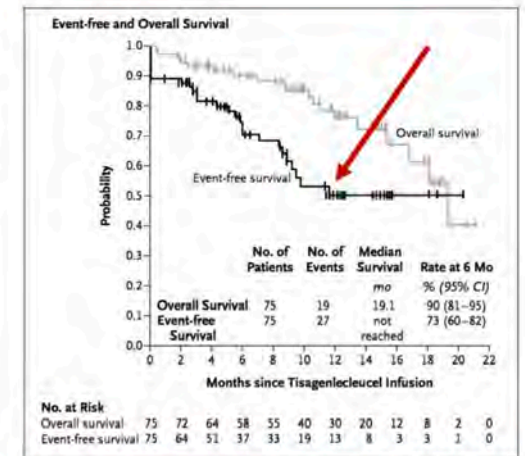
- Retrospective, multicenter study of 420 children and young adults receiving CD19 CAR T-cells
  - 166 (39.5%) with relapse
- Median overall survival (OS):
  - 11.9 months (95% CI: 9.0-17.9 mo)
- 12 month OS: 49.4%
- Salvage options, limited
  - Particularly for CD19<sup>neg</sup> relapse



Lamble A/Shah NN, Blood Adv, 2023

### CD19 CAR T-cells for Long Term Cure

- Need for CAR T-cell persistence and subsequent relapse immunophenotypes differ between B-ALL and mature B-cell lymphomas
- Treatment paradigm for relapsed/refractory B-ALL (pre-CD19 CAR T-cell era):
  - Achieve remission → AlloHSCT
- A fraction of patients will be able to achieve long-term cure with CD19 CAR T-cells alone
  - Avoiding additional short and long-term toxicities from additional chemotherapy and HSCT



Can we improve upon 50% EFS?

Maude SL, et al. NEJM 2018

# Industry appetite for natural killer cells intensifies

**Natural killer cells are attractive as cancer immunotherapy agents because – unlike T cells – they evade immune rejection and do not induce cytokine storms. But capturing their activity in effective therapies remains a work in progress.**

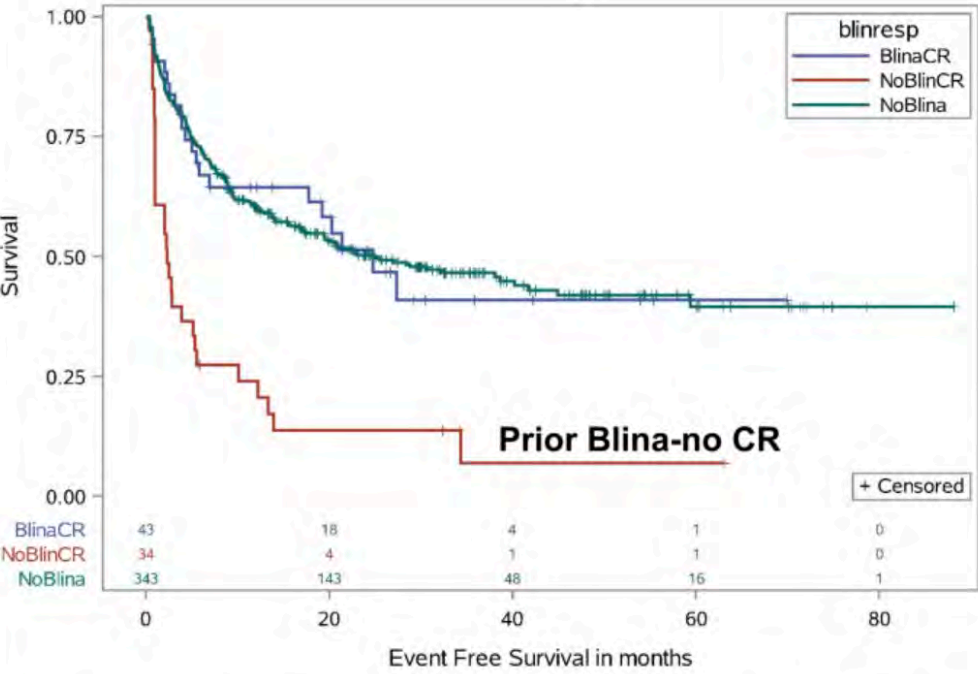
**By Cormac Sheridan**

**I**nterest in natural killer (NK) cells has escalated as large players drum up collaborations to bring NK-driven programs and tools into their portfolios. In December, Sanofi deepened its commitment to antibody-based NK cell engagers, expanding an ongoing partnership with Marseille-based Innate Pharma. A few months earlier, Sanofi



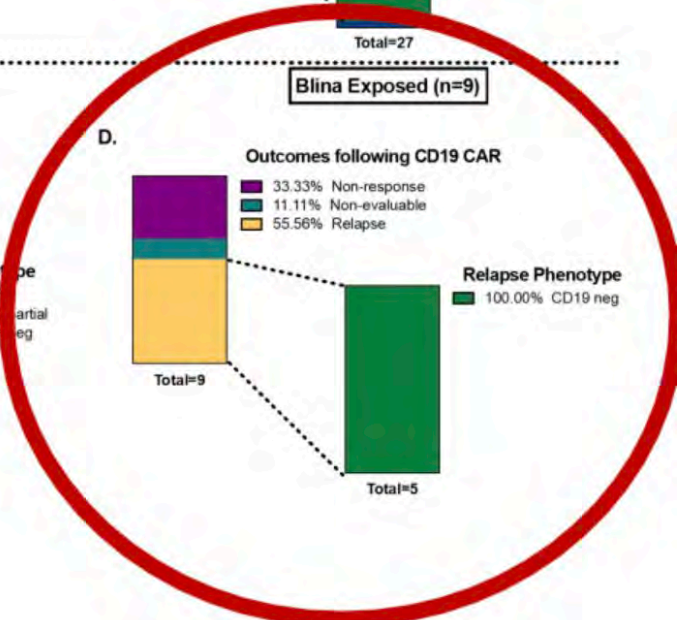
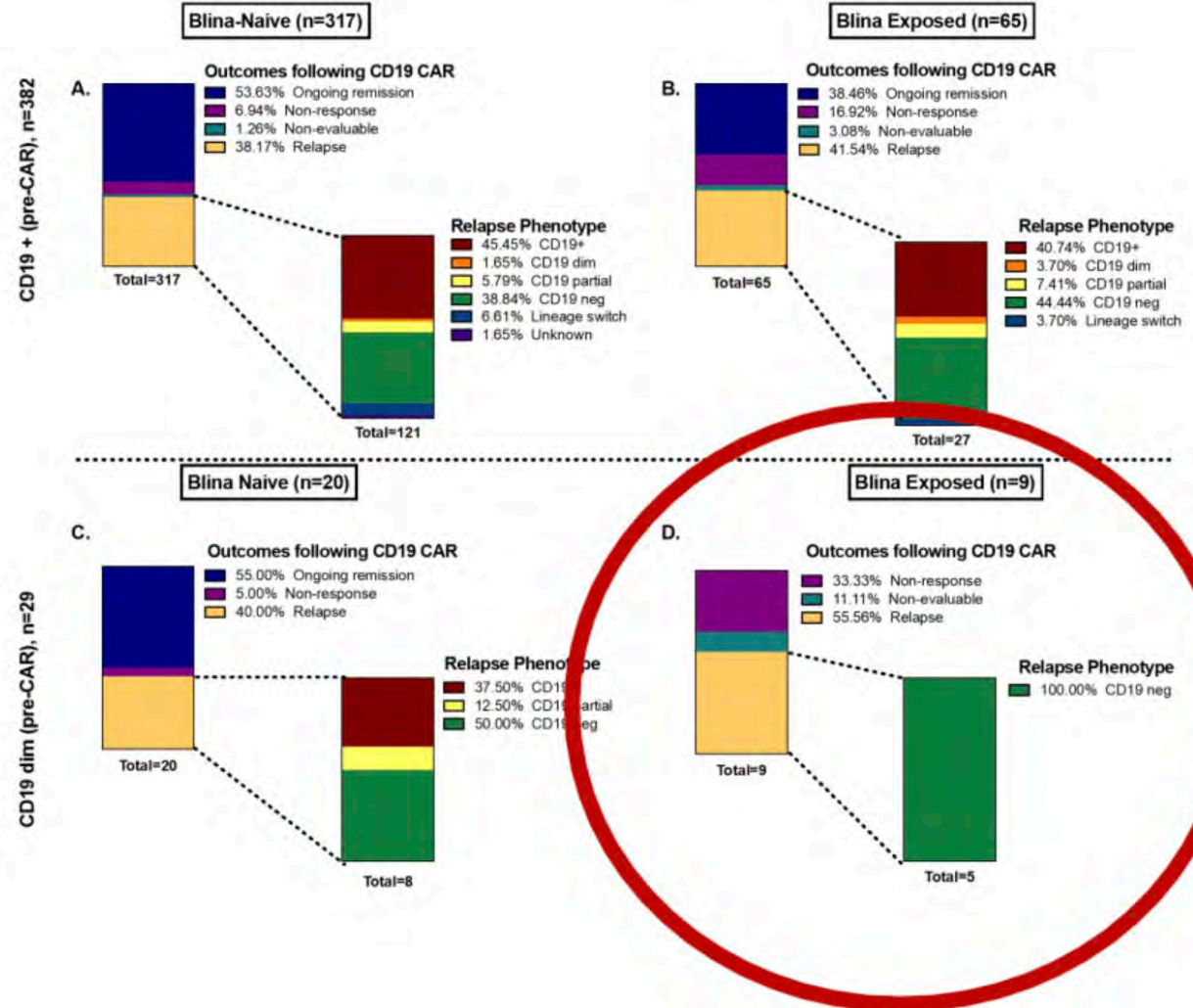
Predicting relapse post CAR T  
An opportunity to improve the outcome

# Pre CAR T-cell Targeted Therapy May Influence Relapse Phenotype



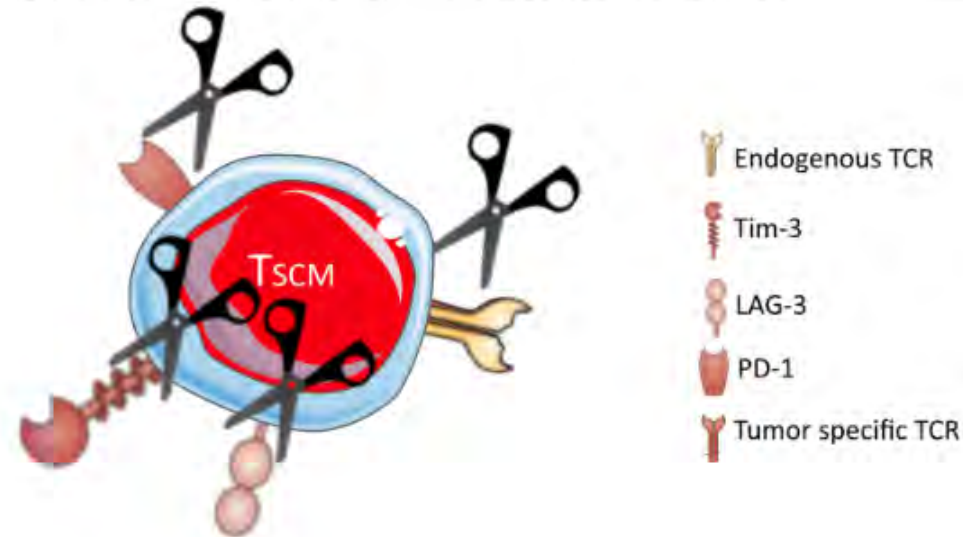
Development of CD19dim disease pre-CAR may influence post CAR T-cell relapse phenotype.

**FURTHER STUDY** is needed



# Adoptive T-cell therapy for cancer: Overcoming T cell exhaustion with genome editing

To **design** and **develop** a panel of **CRISPR/Cas9** molecules targeting genes encoding **inhibitory receptors** in order to enhance the anti-tumor activity of **TCR-edited memory stem T cells**





How do we pivot our research to account for the immune conversation?



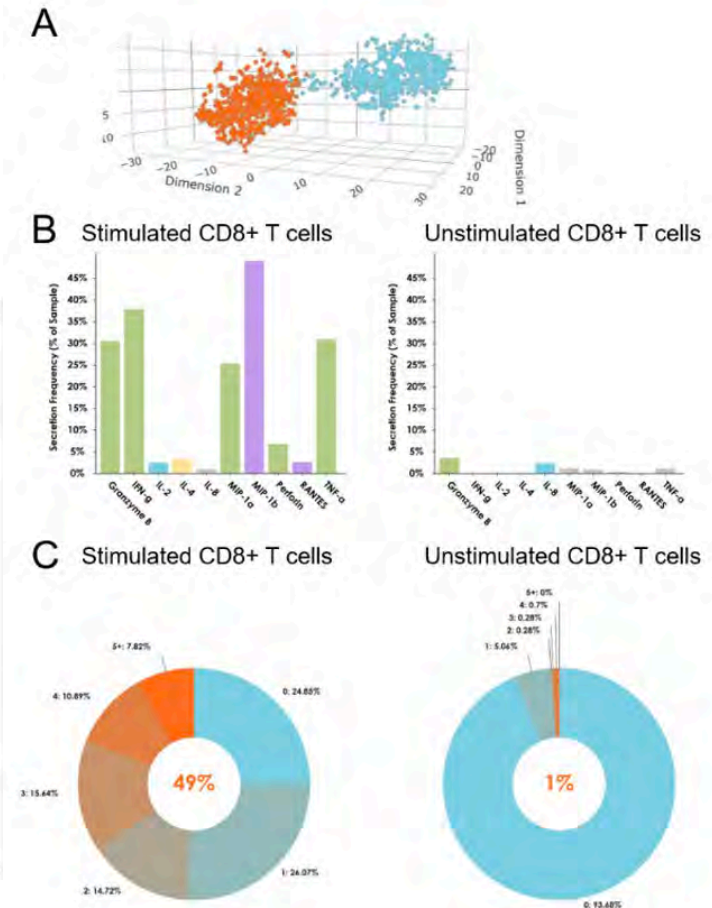
When is the best time to introduce radiation

Never before pheresis? And why  
After pheresis and before infusion?  
How about after infusion?

**Specific Aim 2: Identify changes in circulating T-cell phenotypes and functions before and after CSI or TBI as conditioning for CAR-T cell therapy, and after CAR T**

**Specific Aim 1: Identify changes in circulating inflammatory and immunosuppressive cytokines before at specified time points in both blood and CSF (for those with CNS disease) (before radiation, after radiation and before CAR T, day +7 and +14 of CAR T)**

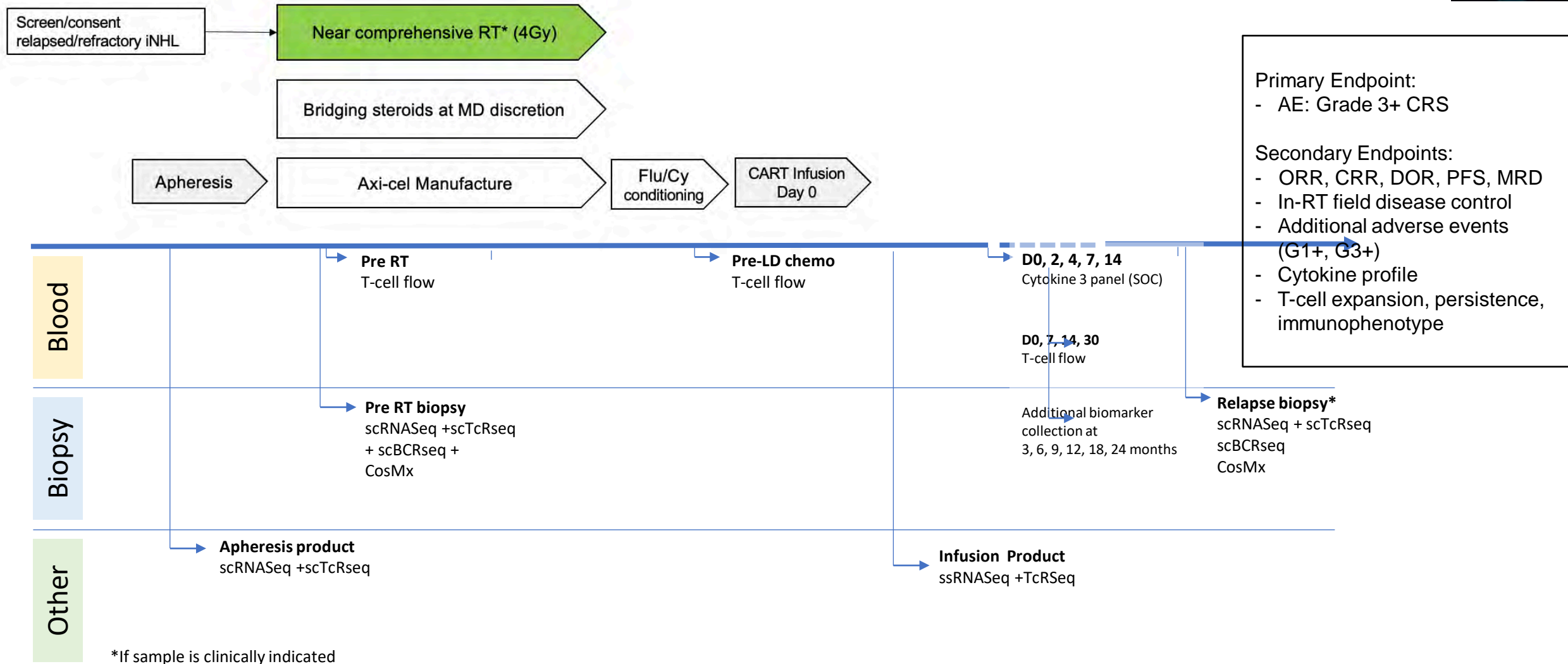
**Figure. Multiplexed immune phenotypic analysis of peripheral blood CD8<sup>+</sup> T cells.** CD8<sup>+</sup> T cells were selected from human peripheral blood mononuclear cells and assayed using the Isolyte adaptive immune response panel. **(A)** tSNE plot between unstimulated (orange) and stimulated (blue) CD8<sup>+</sup> T cells shows distinct phenotypes. **(B)** Cytokines upregulated in stimulated CD8<sup>+</sup> T cells (left panel) compared to unstimulated CD8<sup>+</sup> T cells (right panel). **(C)** Stimulation



# Phase II Study of CAR-T with Bridging RT in Relapsed/Refractory Indolent Non-Hodgkin Lymphoma



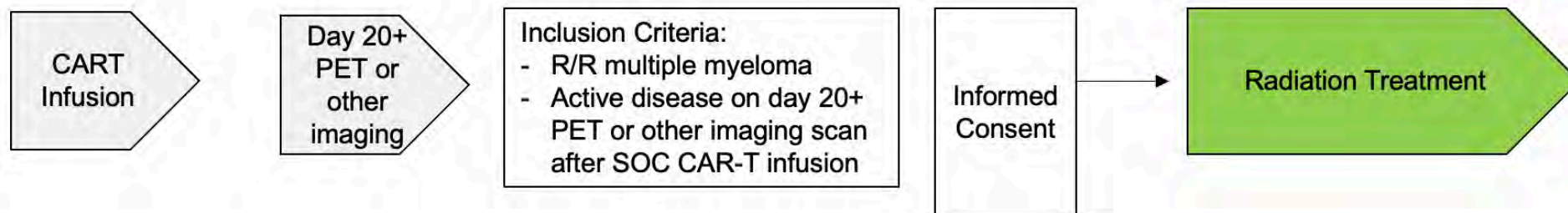
Susan Wu



# Phase II Study of Salvage Radiation Treatment (RT) after B-cell Maturation Antigen (BCMA) Chimeric Antigen Receptor T-cell Therapy (CAR-T) for Relapsed Refractory Multiple Myeloma



Penny Fang



- Study Endpoints:**
- Overall response rate
  - Duration of response among responders
  - Progression free survival
  - Safety
  - Complete response rate
  - Overall survival
  - Local control within RT field

- Biomarker Collection:**
- Optional/Archived biopsy tissue
  - ▲ Blood collection
  - Optional stool collection





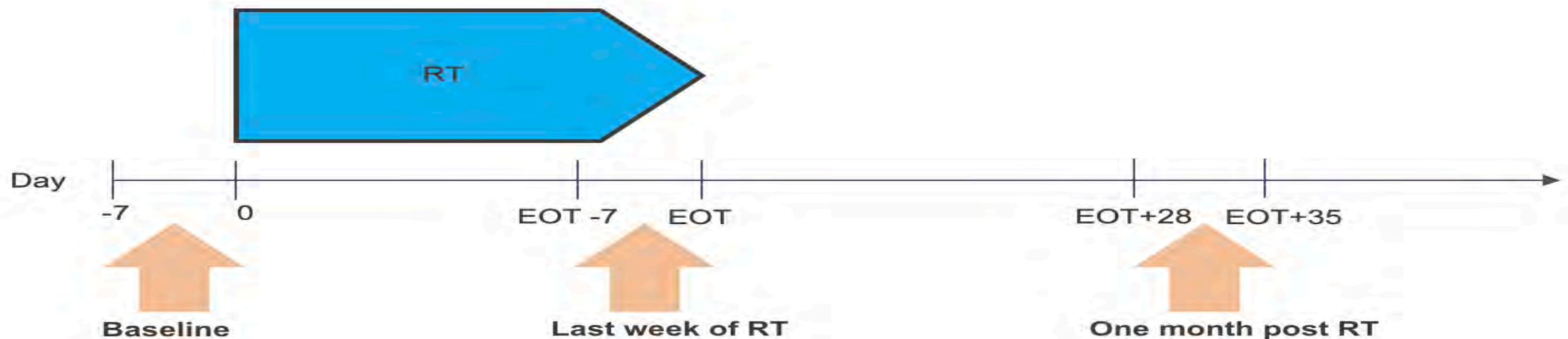
# Umbrella Protocol: 2020-1150 Outcomes after Chimeric Antigen Receptor Therapy (CAR-T) and Radiation Therapy (RT) for Hematologic Malignancies

Primary Objective:

- Study clinical outcomes of patients with hematologic malignancies receiving standard-of-care chimeric antigen receptor therapy (CAR-T) and radiation therapy (RT)

## Secondary Objectives:

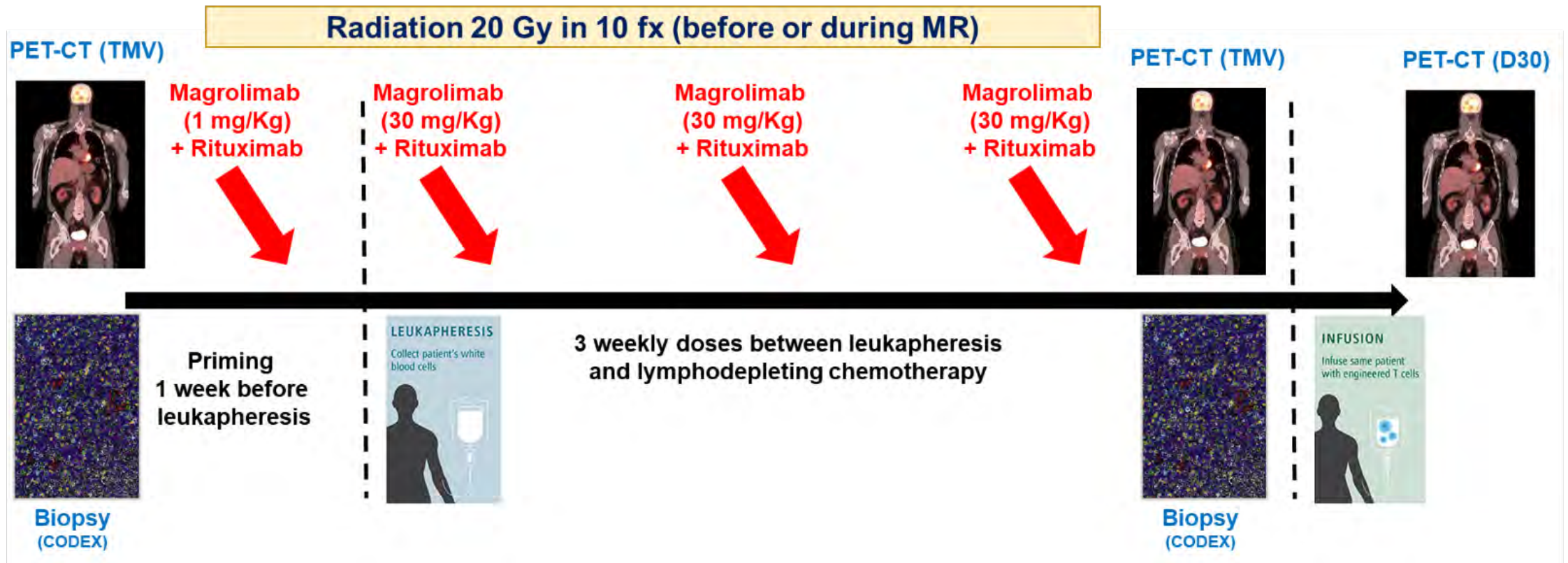
1. To study patient-specific factors and treatment-related factors and outcomes
2. To study the relationship between radiation dose, target, technique, and timing with respect to CAR-T and clinical outcomes in patients
3. To study the relationship between patient-specific factors and treatment-related factors and treatment toxicity



# A Phase II Study of Magrolimab, Rituximab and Radiation as Bridging Strategy Before CAR T-Cell Therapy in Patients with Relapsed or Refractory Large B-cell Lymphoma

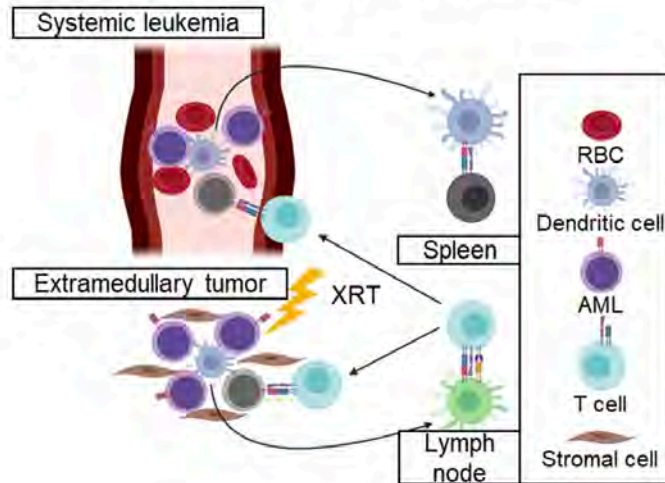


Penny Fang

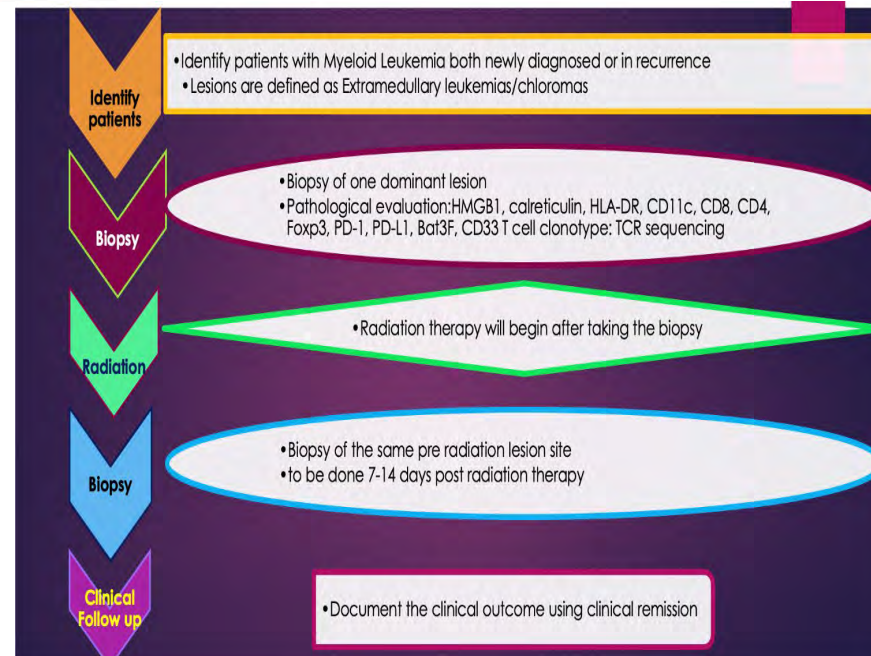
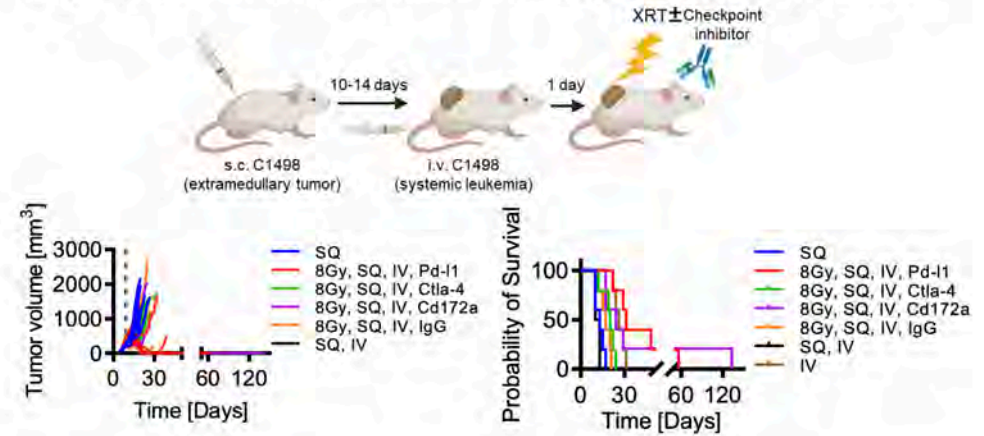


# Radiation and Immunological changes in Chloroma (Extramedullary AML)

## Immune differences between extramedullary and systemic leukemias

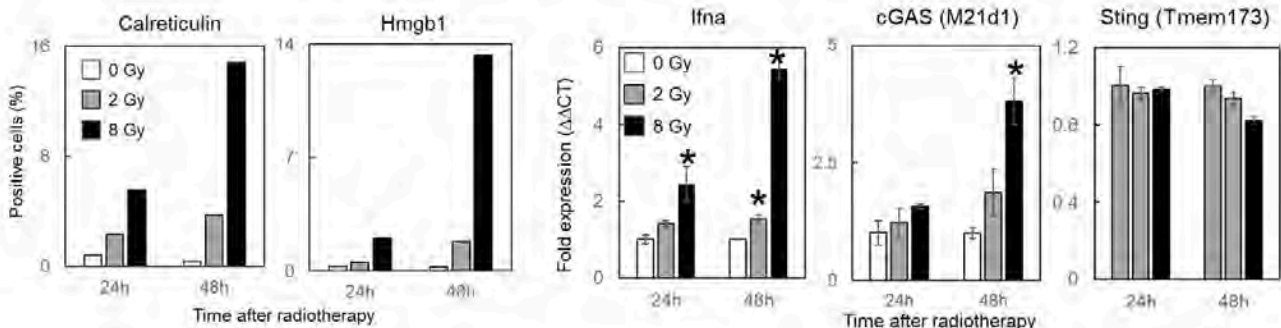


## Radiotherapy with anti-PD-1 or anti-CD47 potentially improves survival in mice with systemic leukemias

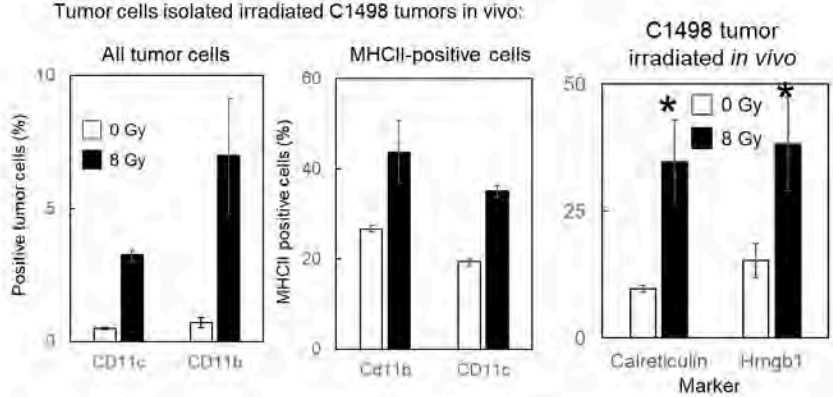


# Radiation and Immunological changes in Chloroma (Extramedullary AML)

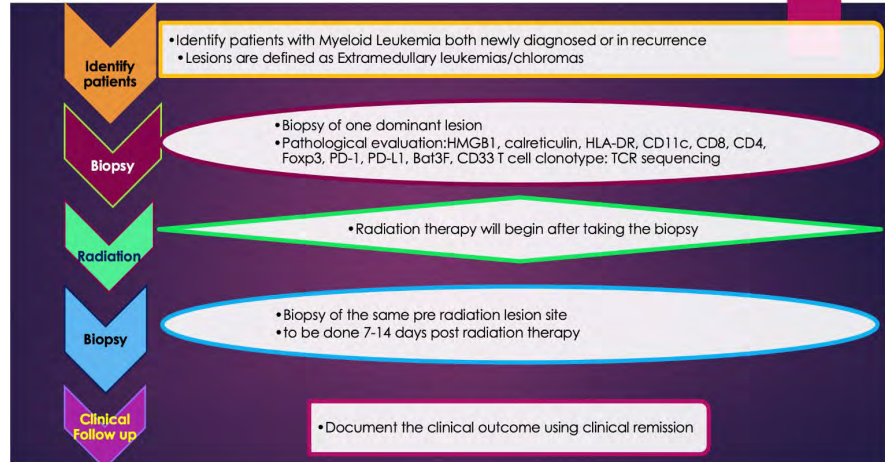
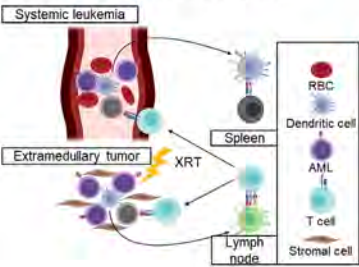
## Radiotherapy increases DAMP and cGAS-STING-IFN pathway expression *in vitro*



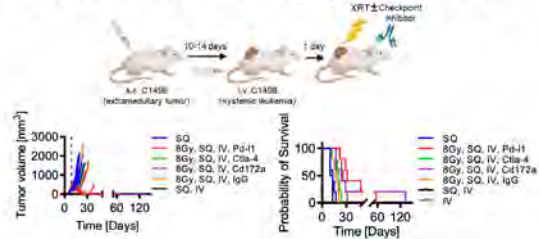
## Radiotherapy increases macrophage infiltration and DAMP expression *in vivo*



## Immune differences between extramedullary and systemic leukemias



## Radiotherapy with anti-PD-1 or anti-CD47 potentially improves survival in mice with systemic leukemias



Mechanism of Radiation priming

Correct timing to introduce Radiation

How does Radiation affect the CAR T cell therapy persistence fitness

How does it affect the outcome

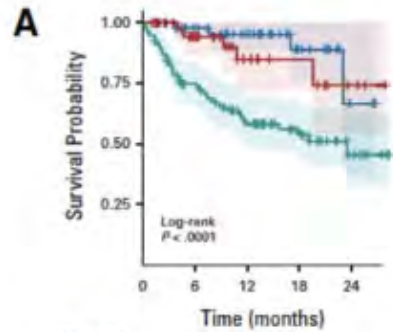
How to use the aforementioned as a steppingstone towards bi-specifics

*We have a lot of work to do*

*The answer is through  
Translational Studies*

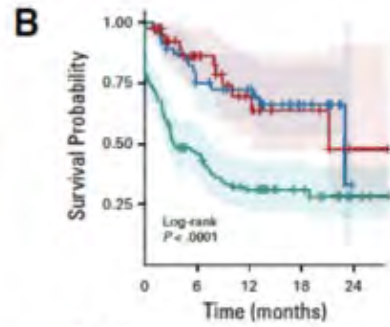
# Debulking

## Disease Burden Impacts Response and Toxicity



No. at risk:

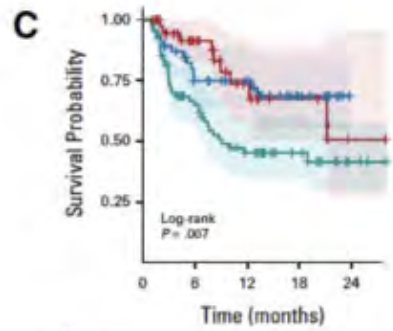
Strata	46	38	28	12	2
—	41	28	14	8	3
—	93	62	40	24	7



No. at risk:

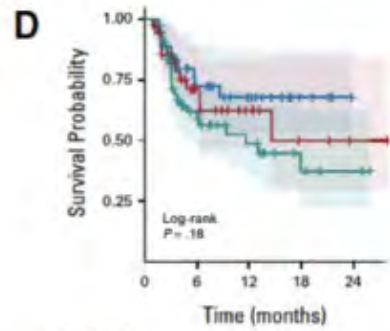
Strata	46	30	25	10	0
—	41	25	12	5	1
—	93	38	22	13	3

— No detectable disease  
— Low-disease burden  
— High-disease burden



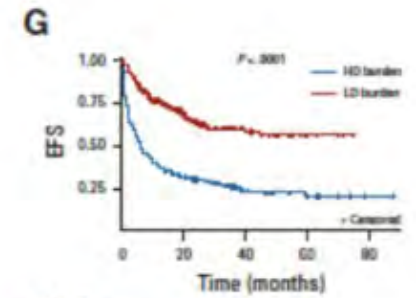
No. at risk:

Strata	46	30	25	10	0
—	41	25	12	5	1
—	93	38	22	13	3



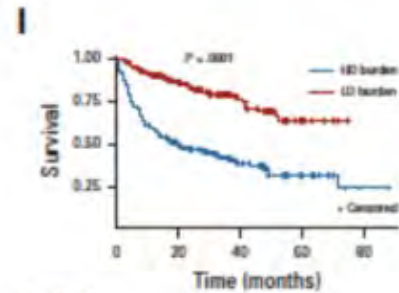
No. at risk:

Strata	37	19	12	4	0
—	32	14	8	3	0
—	74	43	20	7	0



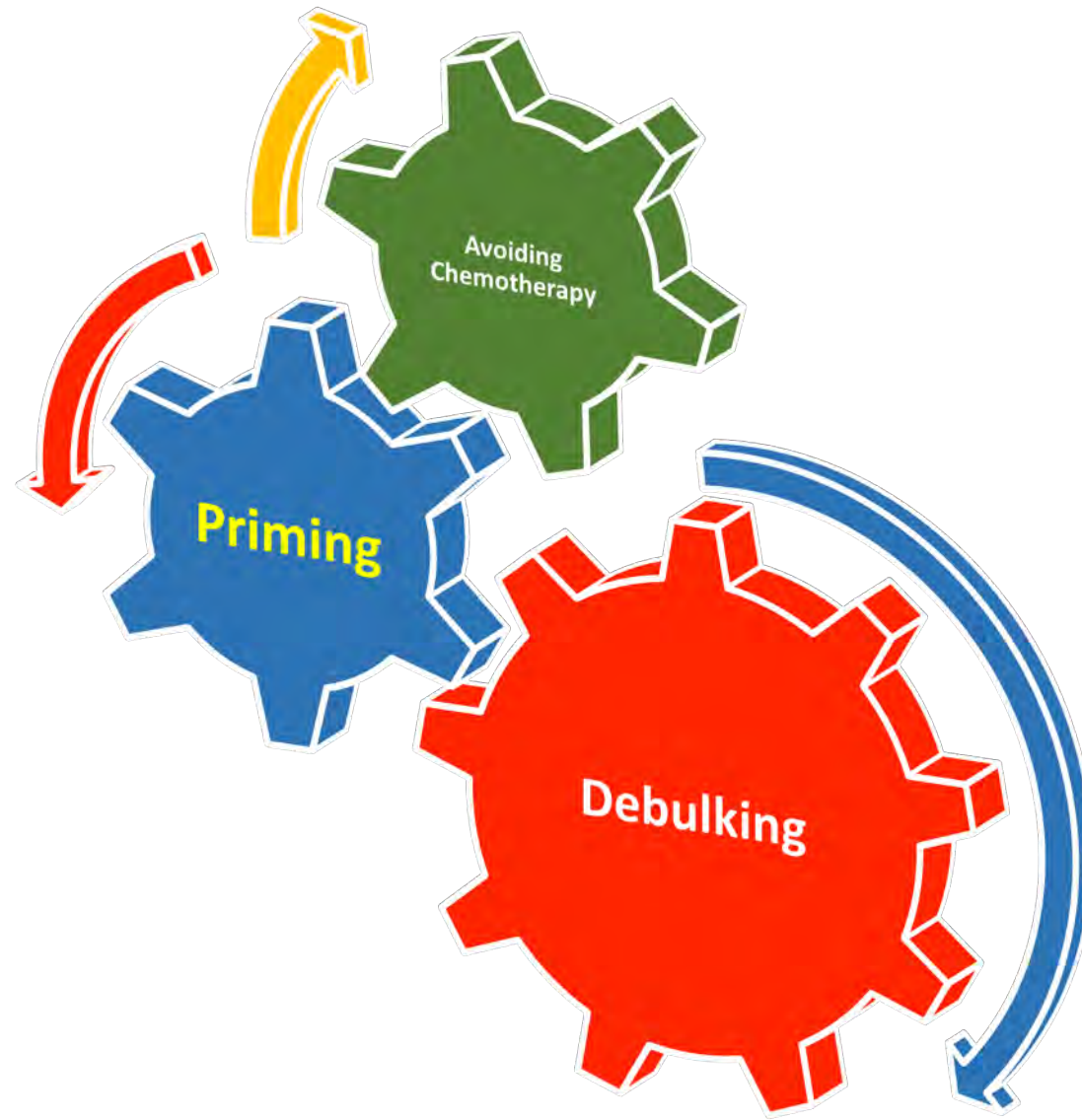
No. at risk:

HD burden	303	40	25	9	1
LD burden	217	95	22	9	6

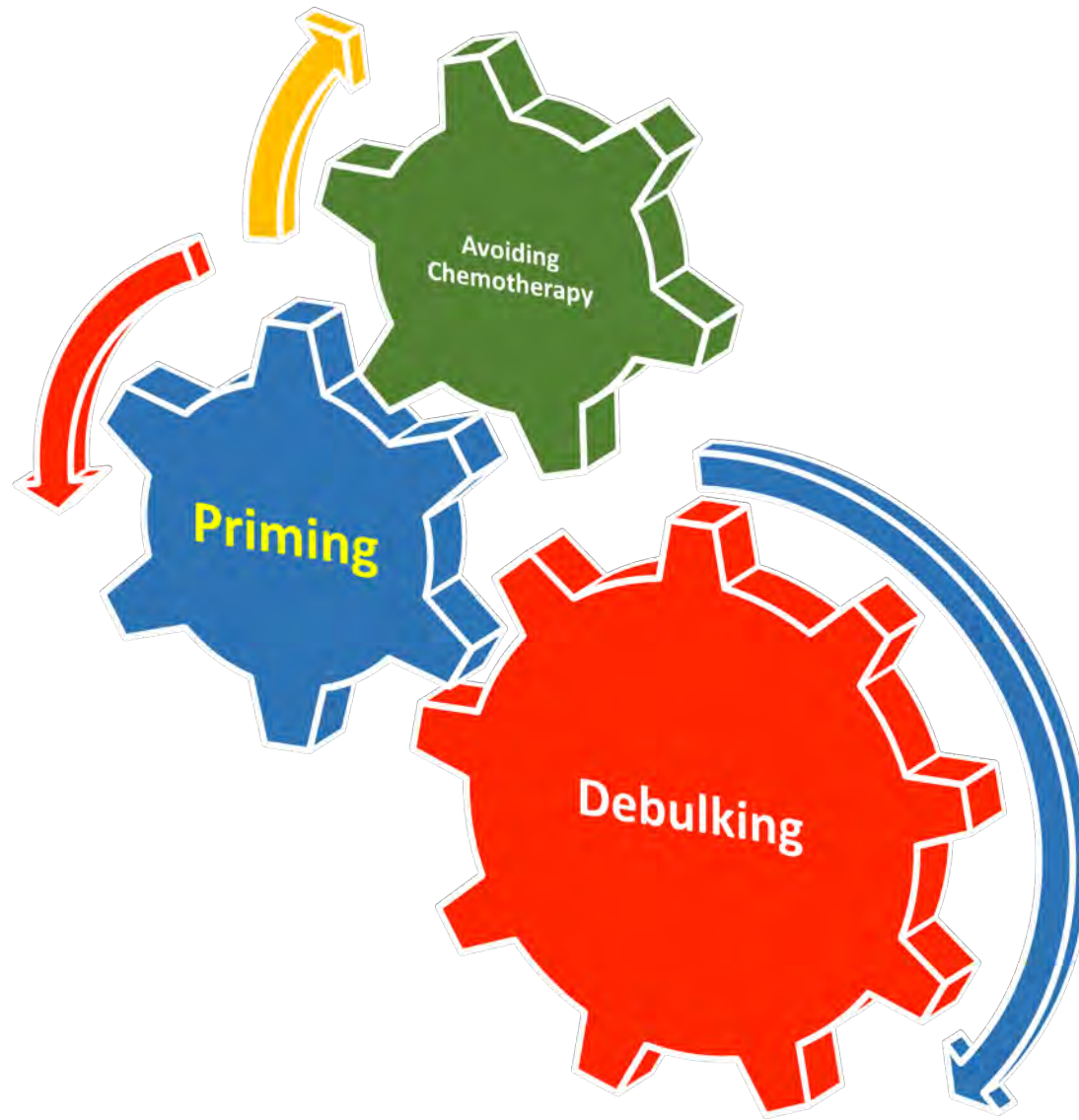


No. at risk:

HD burden	303	95	38	17	1
LD burden	217	128	65	12	1



**Priming**

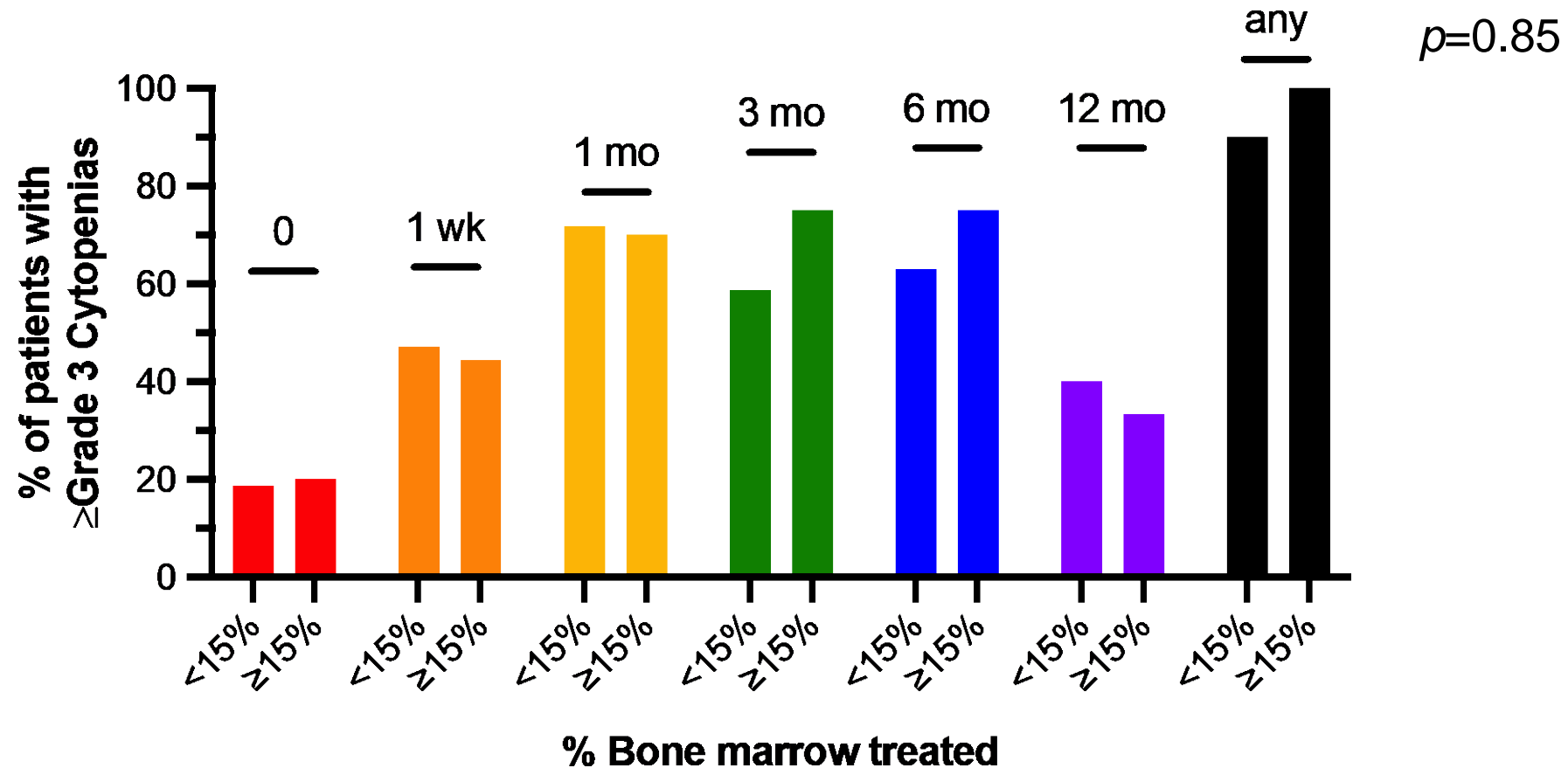




# ≥15% BM treated with bridging RT does not correlate with ≥Grade 3 cytopenias at any timepoint

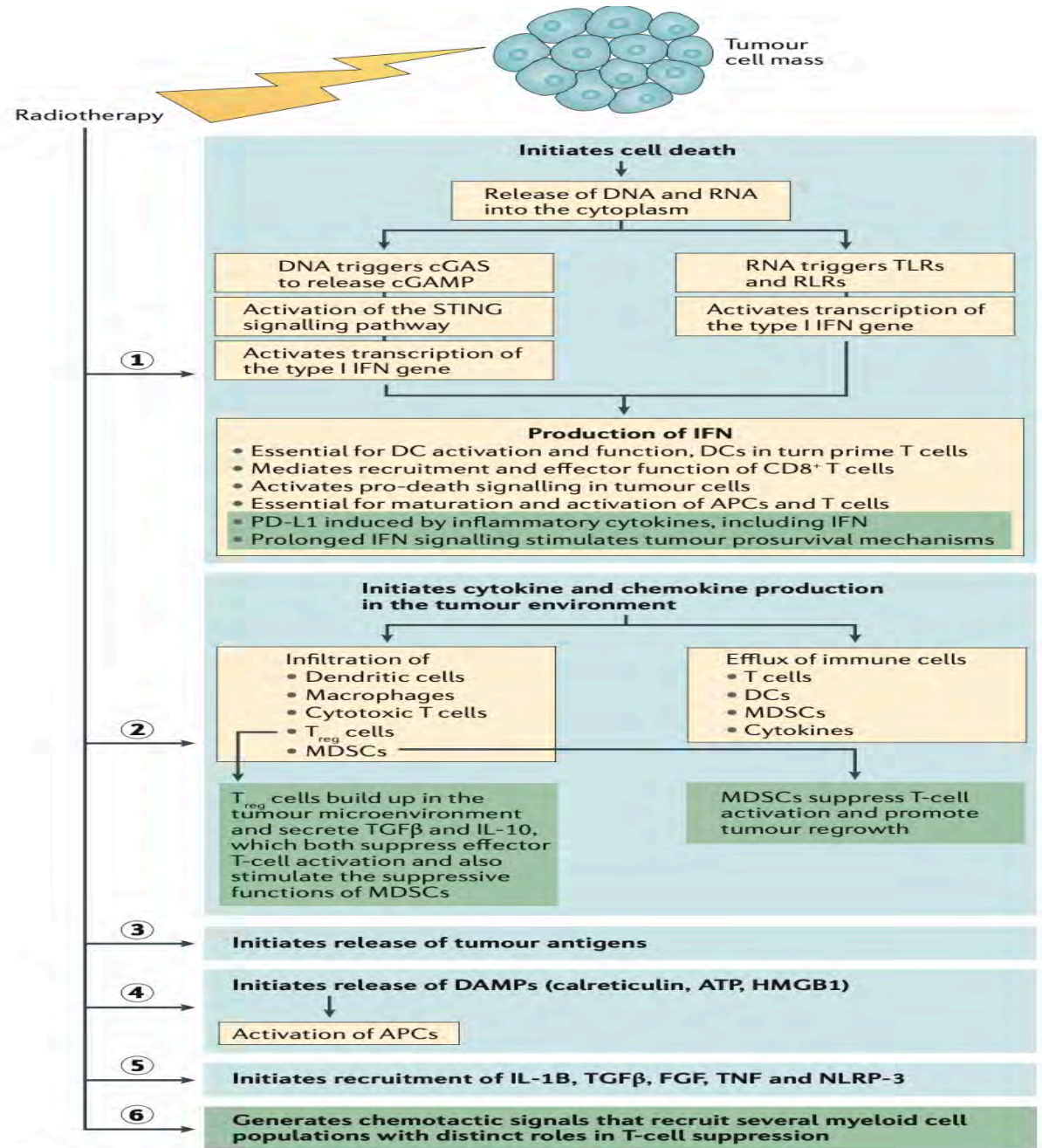


Gohar Manzar



# Radiotherapy and immunotherapy: a beneficial liaison?

Ralph R. Weichselbaum<sup>1</sup>, Hua Liang<sup>1</sup>, Liufu Deng<sup>1</sup> and Yang-Xin Fu<sup>2</sup>



# Ionizing radiation modulates the phenotype and function of human CD4+ induced regulatory T cells



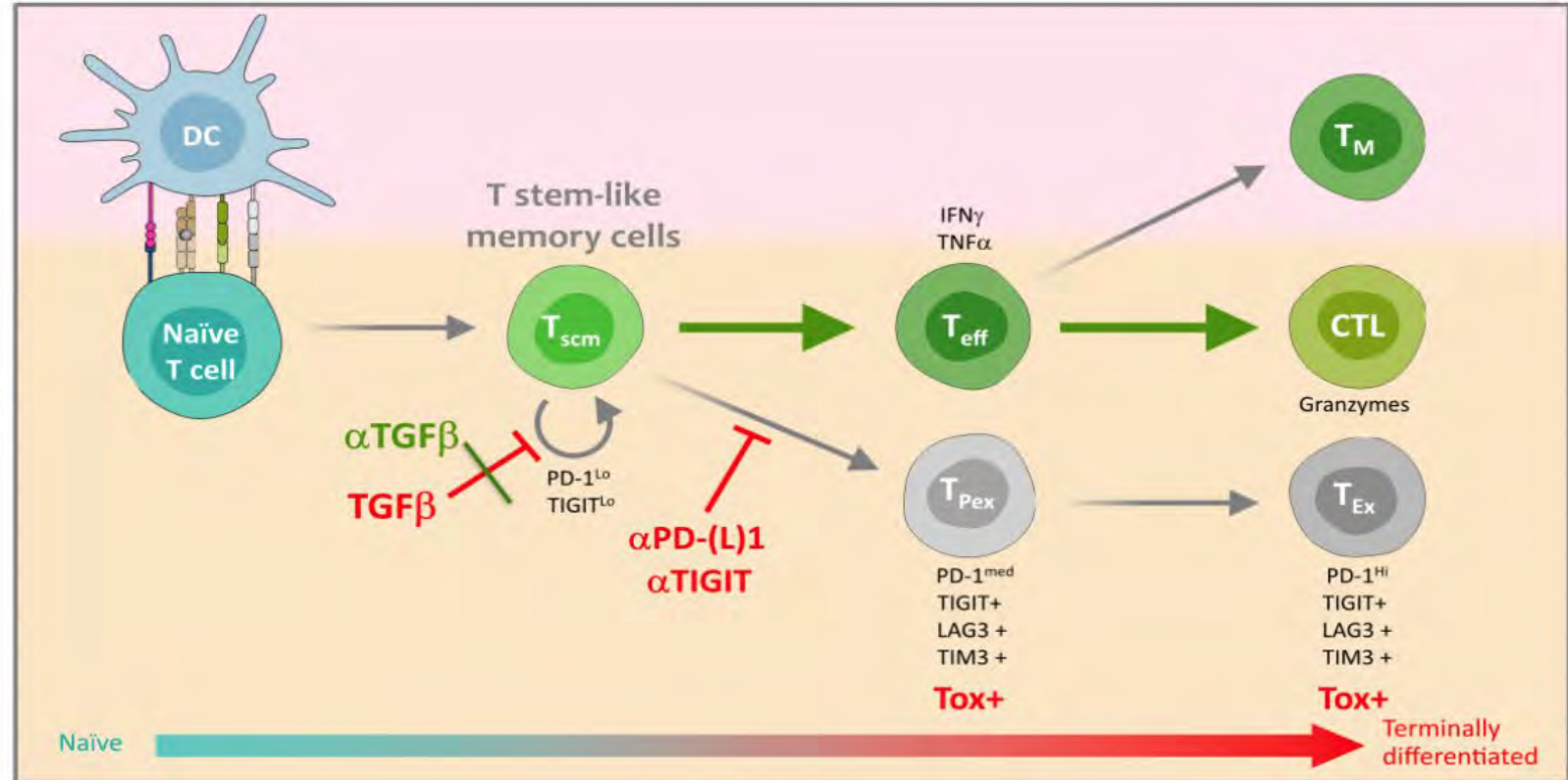
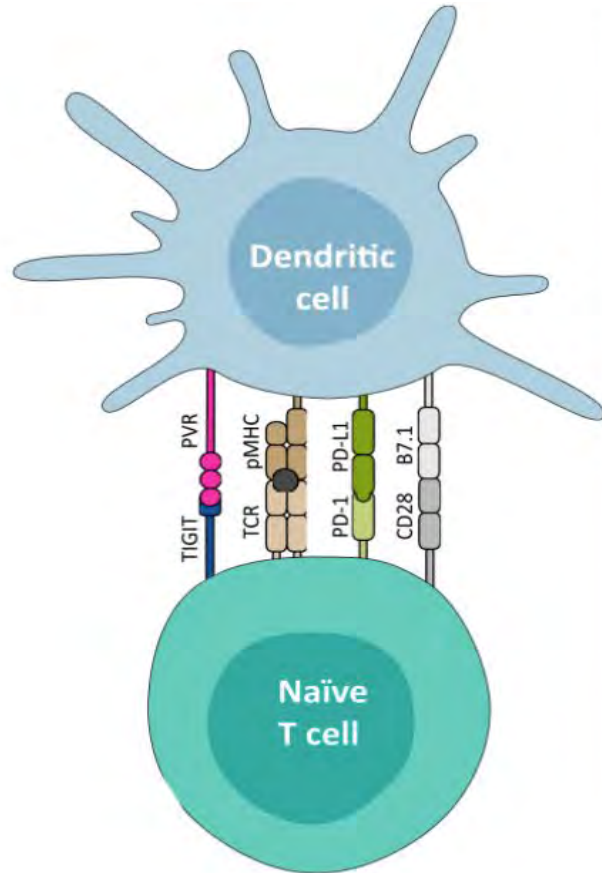
Samantha S. Beauford, Anita Kumari and Charlie Garnett-Benson\*

**Conclusions:** Our findings demonstrate that while human T<sub>REG</sub> cells are more resistant to radiation-induced death, treatment causes downregulation of Foxp3 expression, as well as modulation in the expression of T<sub>REG</sub> signature molecules associated with suppressive activity. Functionally, irradiated TGF- $\beta$ 1-induced T<sub>REGS</sub> were less effective at inhibiting CD8+ T cell proliferation. These data suggest that doses of radiotherapy in the hypofractionated range could be utilized to effectively target and reduce T<sub>REG</sub> activity, particularly when used in combination with cancer immunotherapies.

*Opportunity for Radiation*

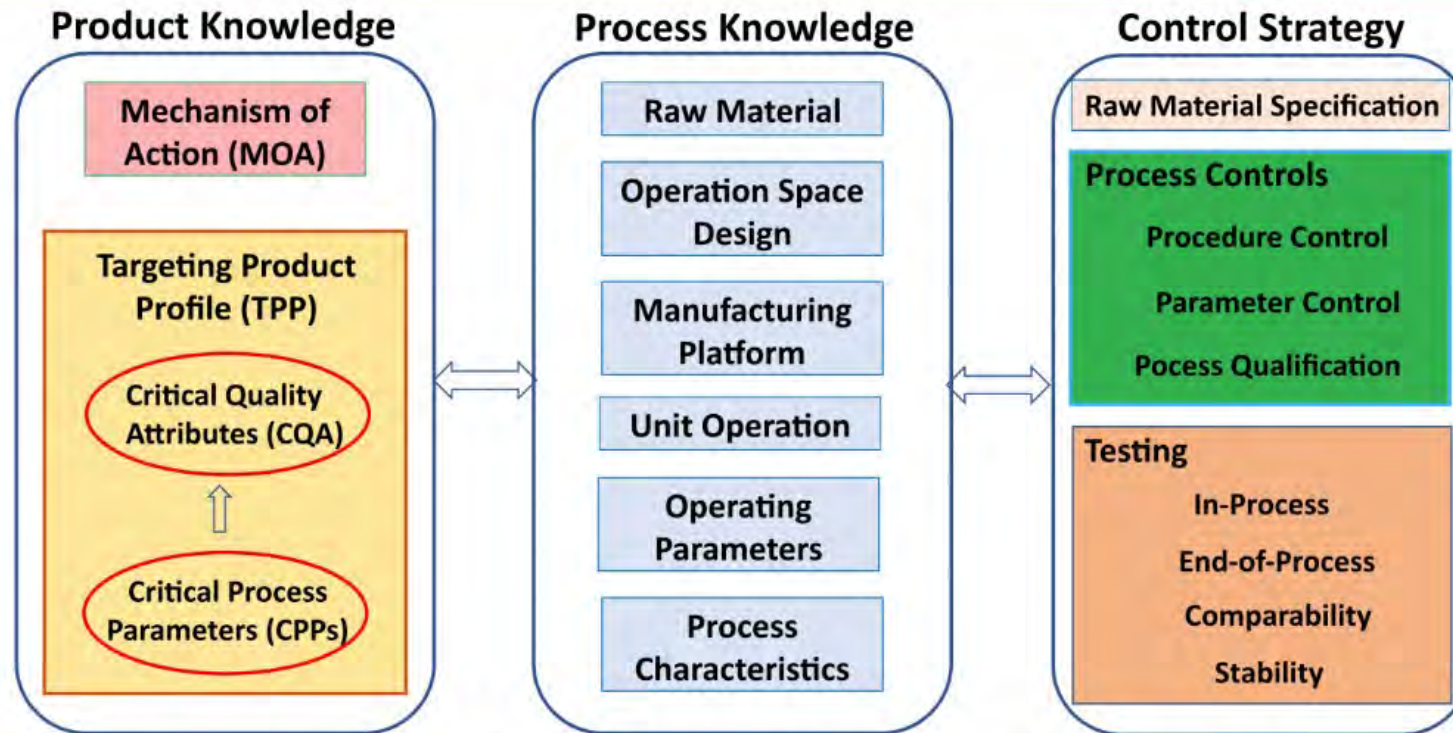
Combined PD-L1/PD-1 and TIGIT blockade re-directs the differentiation of activated T cells to  $T_{\text{eff}}/T_{\text{mem}}$  rather than  $T_{\text{ex}}$

TGF $\beta$  blockade may expand Tscm compartment, further enhancing differentiation of Teff/Tm cells



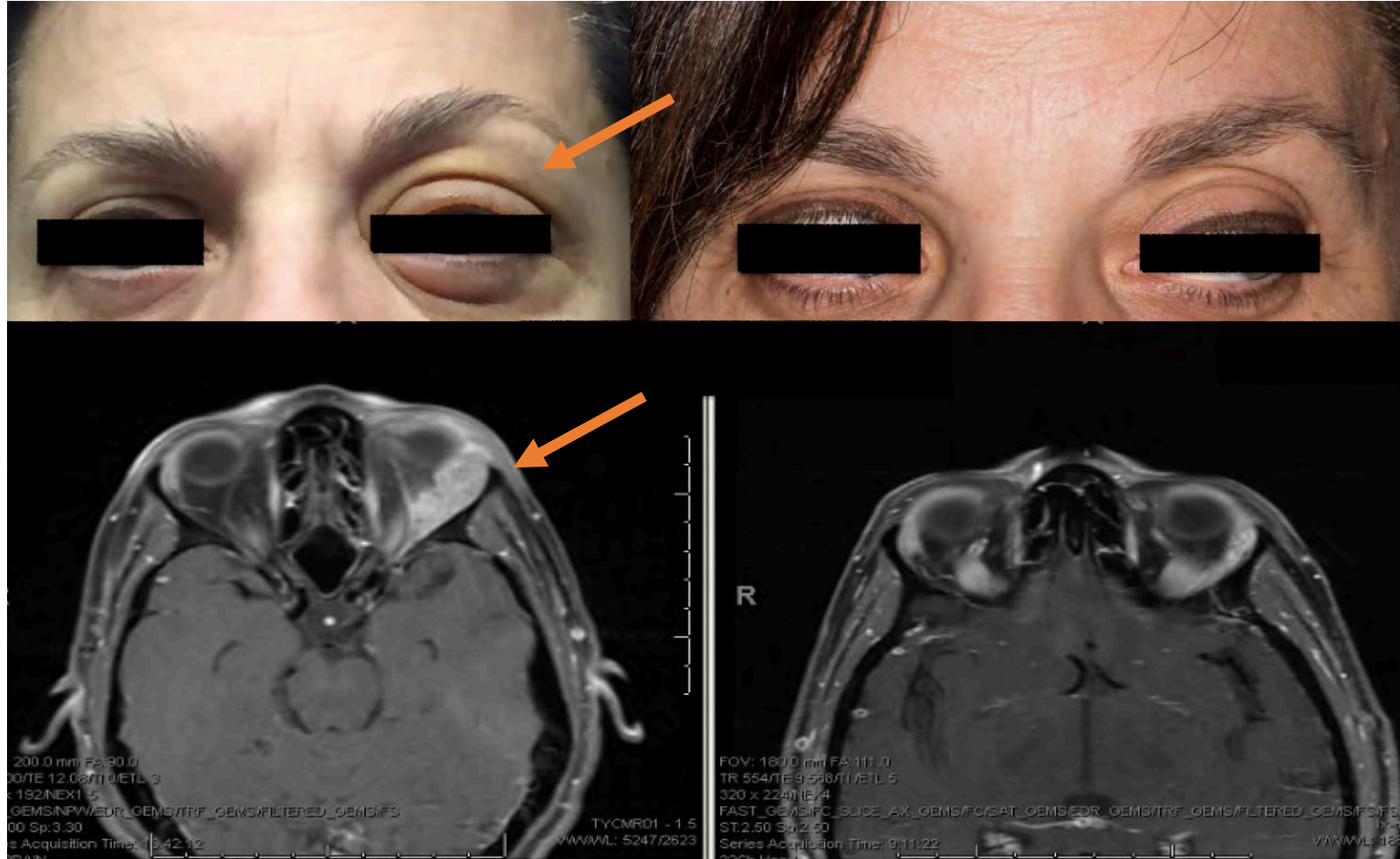
Castiglioni et al (2023) in revision  
Hu et al (2022) JEM  
Ma et al (2022) JEM

## Production process design and development for cell therapy products



Production optimization and establishment of proper process con

As opposed to solid tumors  
Activating the immune microenvironment has been clinically successful  
Boom Boom= 2 Gy x 2

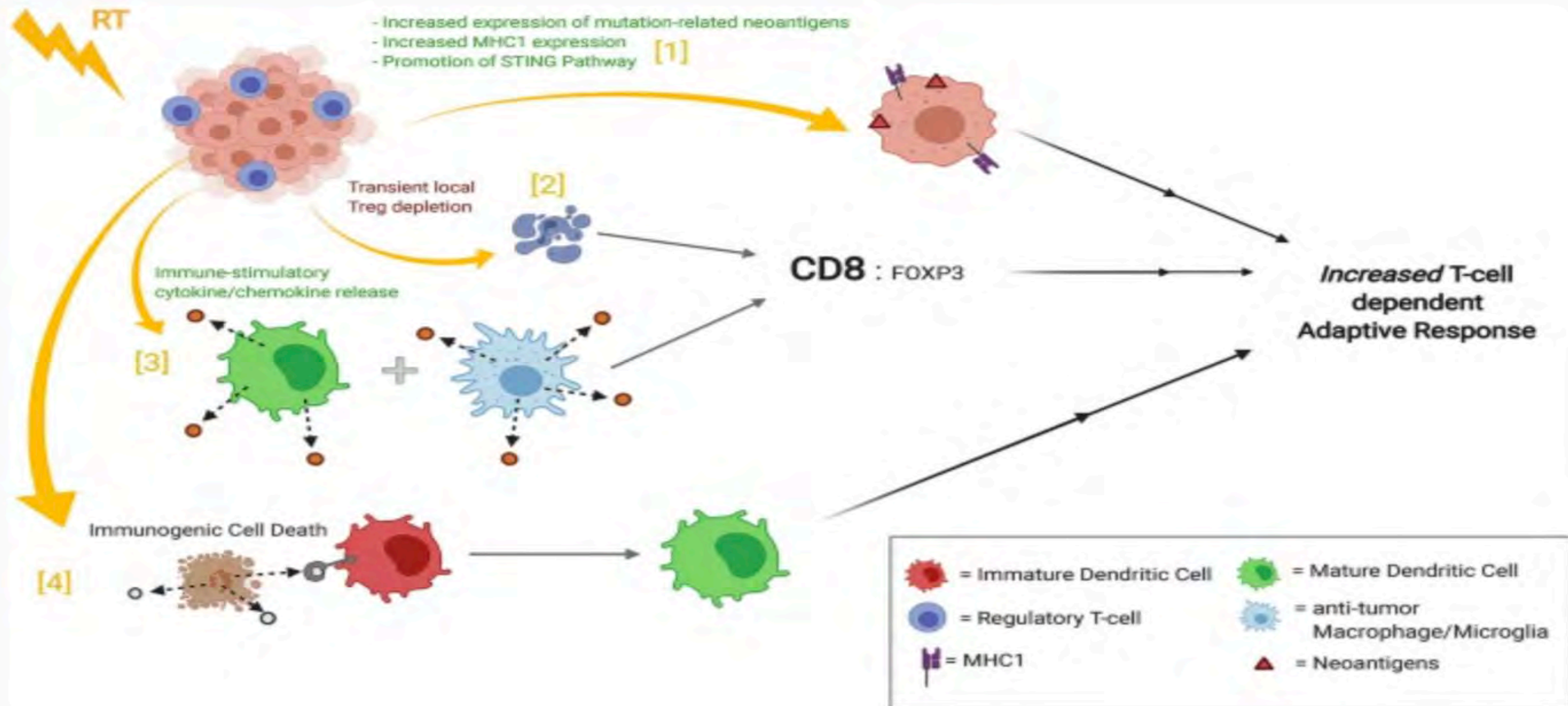


# Rapid manufacturing (24-48 hrs)

Protocol	Target	Indication	Process	Length	Cell dose (CAR+ cells)	Response	CRS ICANs		
NCT04638270	CD19	ALL	Fast CART	1 day	5 x 10 <sup>4</sup> CAR+ T/kg (n=3) 1 x 10 <sup>5</sup> CAR+ T/kg (n=4) 1.5 x 10 <sup>5</sup> CAR+ T/kg (n=3)	100% CR 90% MRD-	90%	Zhang et al	ASH 2019
NCT04129099	CD19/CD22	ALL	Fast CART Gracell	1 day	6.0 x 10 <sup>4</sup> /kg (n=2) 1.0-1.5 x 10 <sup>5</sup> /kg (n=7) 2.25 x 10 <sup>5</sup> /kg (n=1)	90% CR	60%	Yang et al	ASH 2020
NCT04318327 PHE885	BCMA (41BBz)	Myeloma	T- Charge™ Novartis	< 2 days	2.5 x 10 <sup>6</sup> (n=4) 5 x 10 <sup>6</sup> (n=10) 14.3 x 10 <sup>6</sup> (n=1)	33% sCR, 93% ORR		Sperling et al	ASH 2021
NCT05172596 PHE885 Phase II	BCMA (41BBz)	Myeloma	T- Charge™ Novartis	< 2 days	5.0 x 10 <sup>6</sup>	N/A		Munshi et al	EHA, 2022
NCT03960840 YTB323	CD19 (41BBz)	DLBCL	T- Charge™ Novartis	2 days	1-2.5 x 10 <sup>6</sup> (n=4) 5-12.5 x 10 <sup>6</sup> (n=10) 25-40 x 10 <sup>6</sup> (n=1)	73% (3 mo)	27%	Flinn et al	ASH 2021
					1-2.5 x 10 <sup>6</sup> (n=4) 5-12.5 x 10 <sup>6</sup> (n=28) 25 x 10 <sup>6</sup> (n=7) 40 x 10 <sup>6</sup> (n=6)	@DL2, 63% (12/19, 3 mo), 69% (11/16, 6 mo)	33% 11%	Barba et al.	ASH 2022

Rapid manufacturing of non-activated potent CAR T cells. Milone et al. Nat Biomed engineering. 2022  
24 hrs, Serum Starvation, IL-7/IL-15, Deoxynucleosides

**Fig. 1**

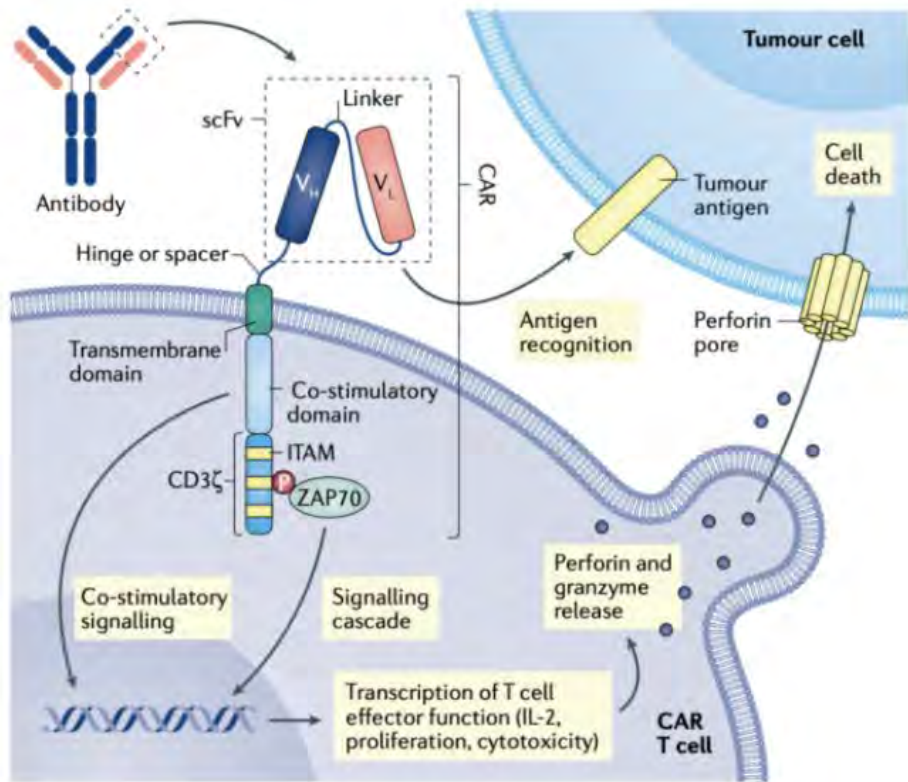


**Strategies that improve antigen presentation:**

- Induce epitope spreading,
- Perpetuate existing antitumor T cell responses
- Combat tumor antigen escape.

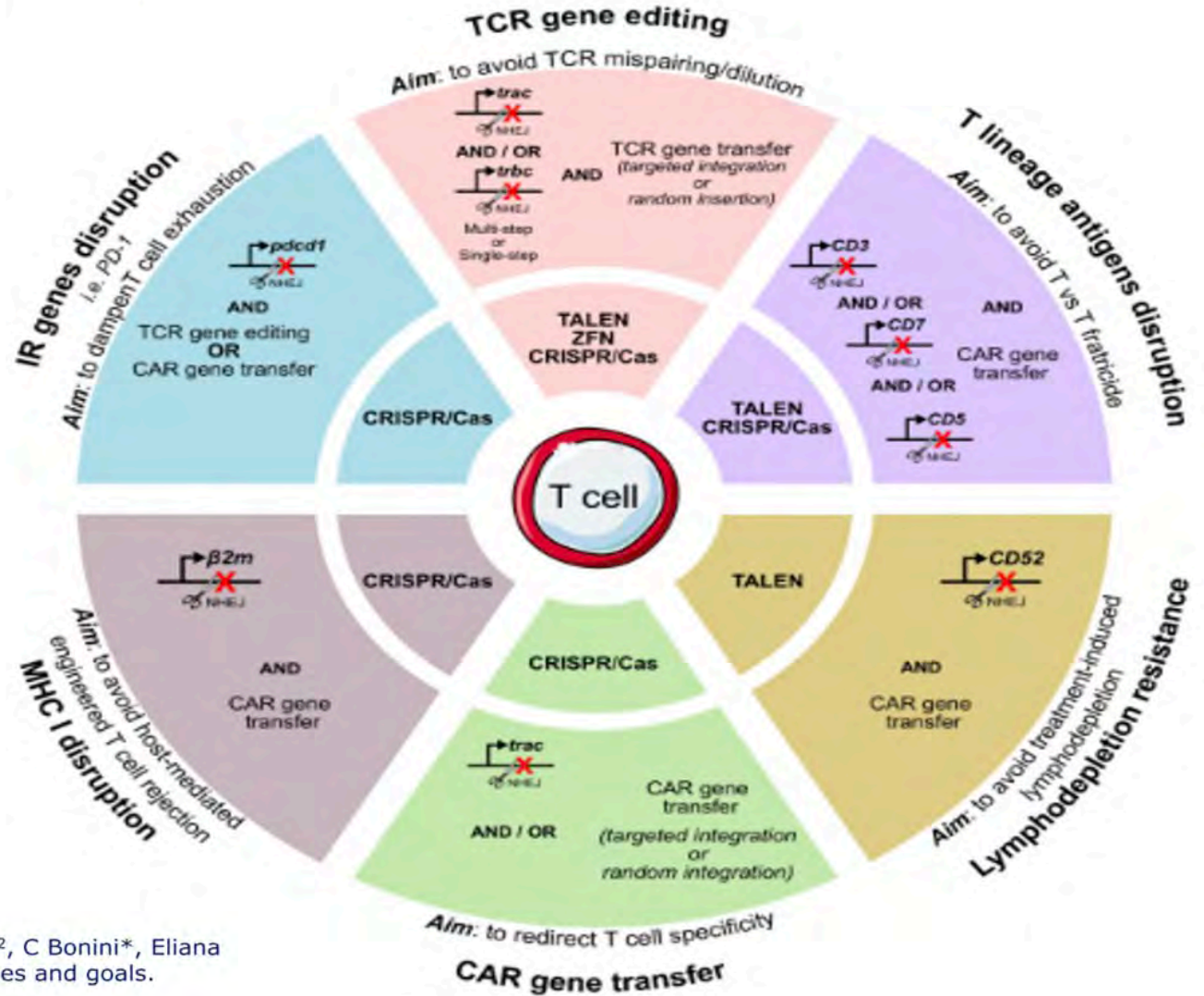


# What are the important mechanisms behind CAR T cells?



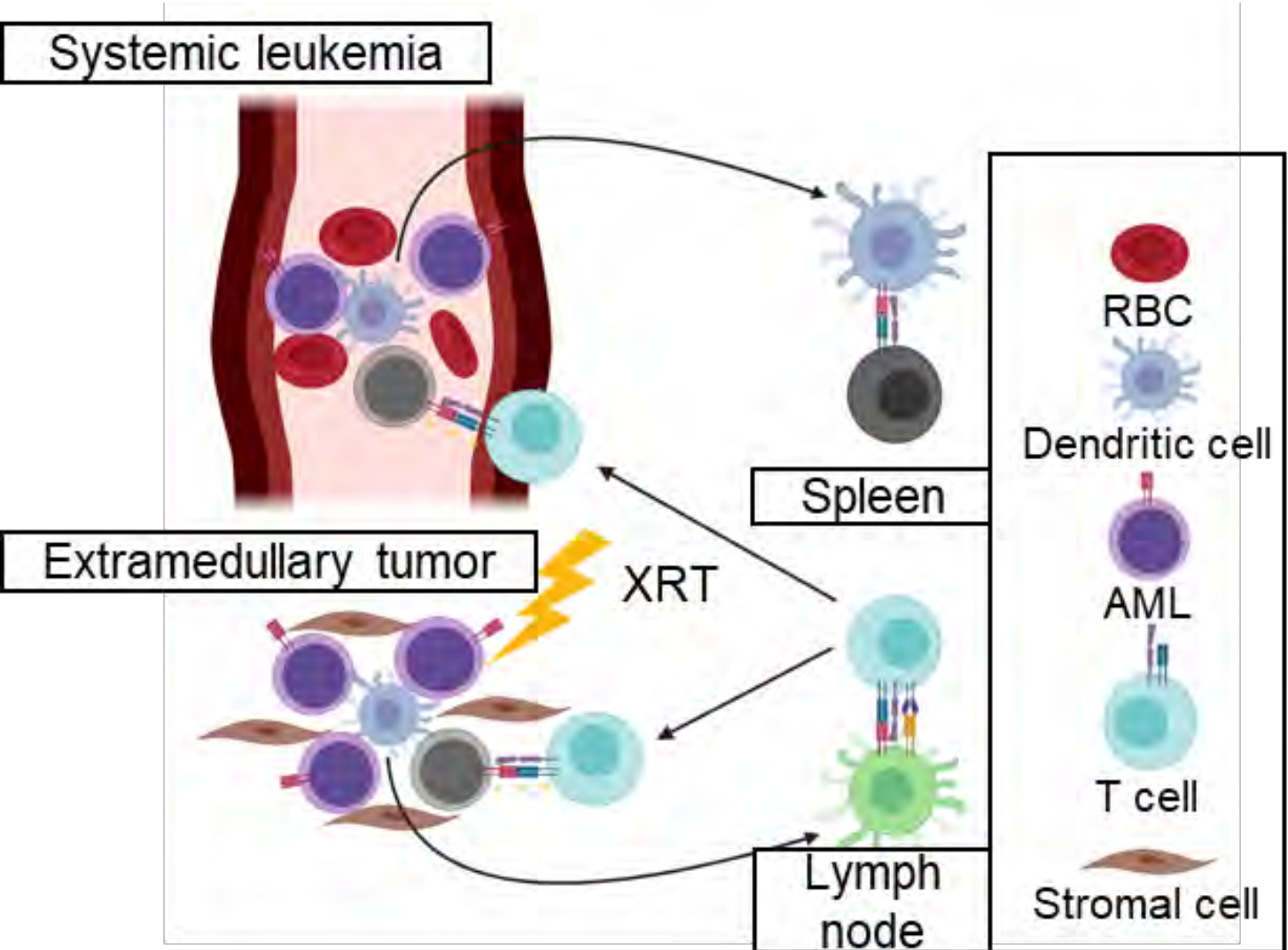
- Induce tumor lysis / trigger apoptosis
- Produce cytokines
- Recruit bystanders
- Proliferate
- Persist / form memory

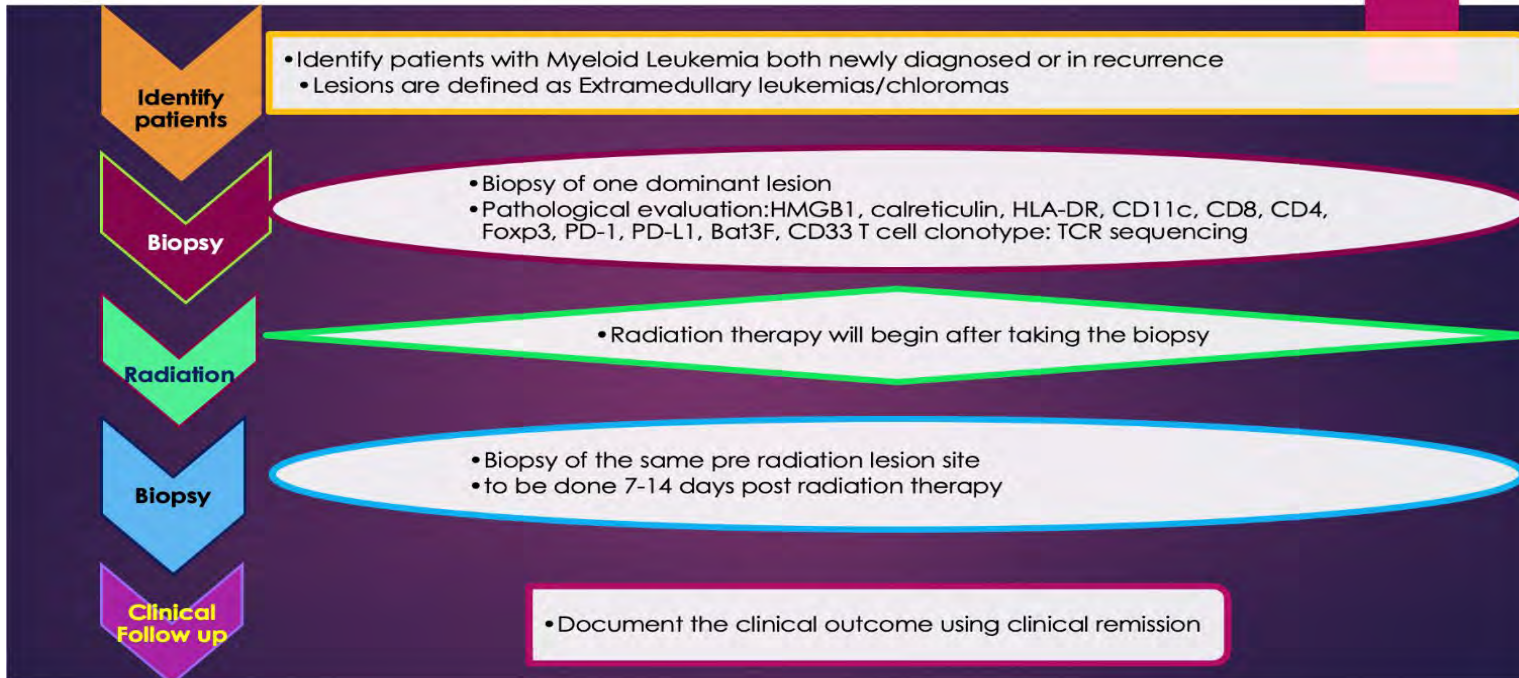
# Genome editing at service of ACT



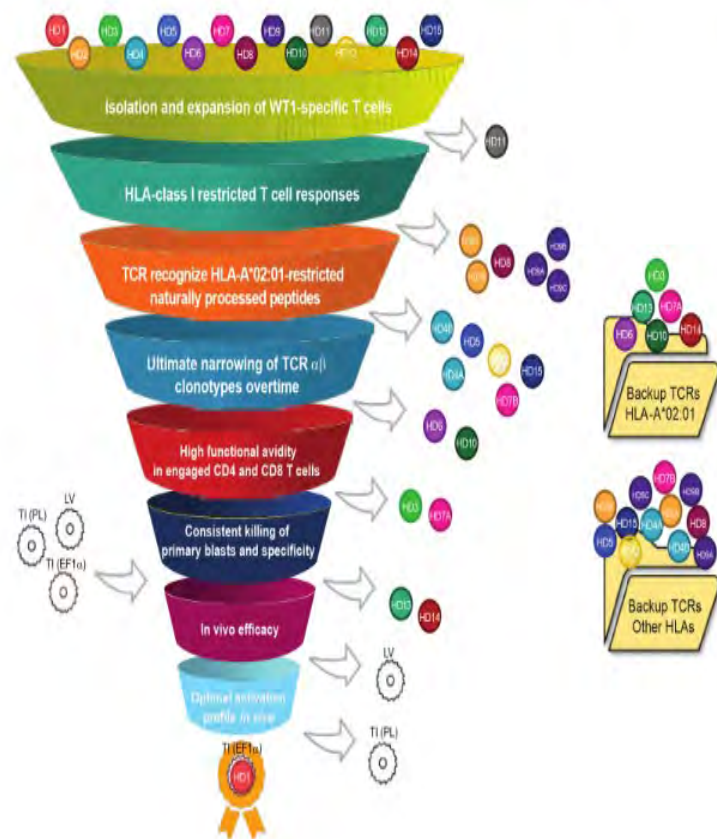
F Manfredi, BC Cianciotti, A Potenza, E Tassi, M Noviello, A Biondi, F Ciceri<sup>1,2</sup>, C Bonini\*, Eliana Ruggiero\* TCR redirected T cells for cancer treatment: achievements, hurdles and goals. *Frontiers in Immunol.* 2021

# Immune differences between extramedullary and systemic leukemias





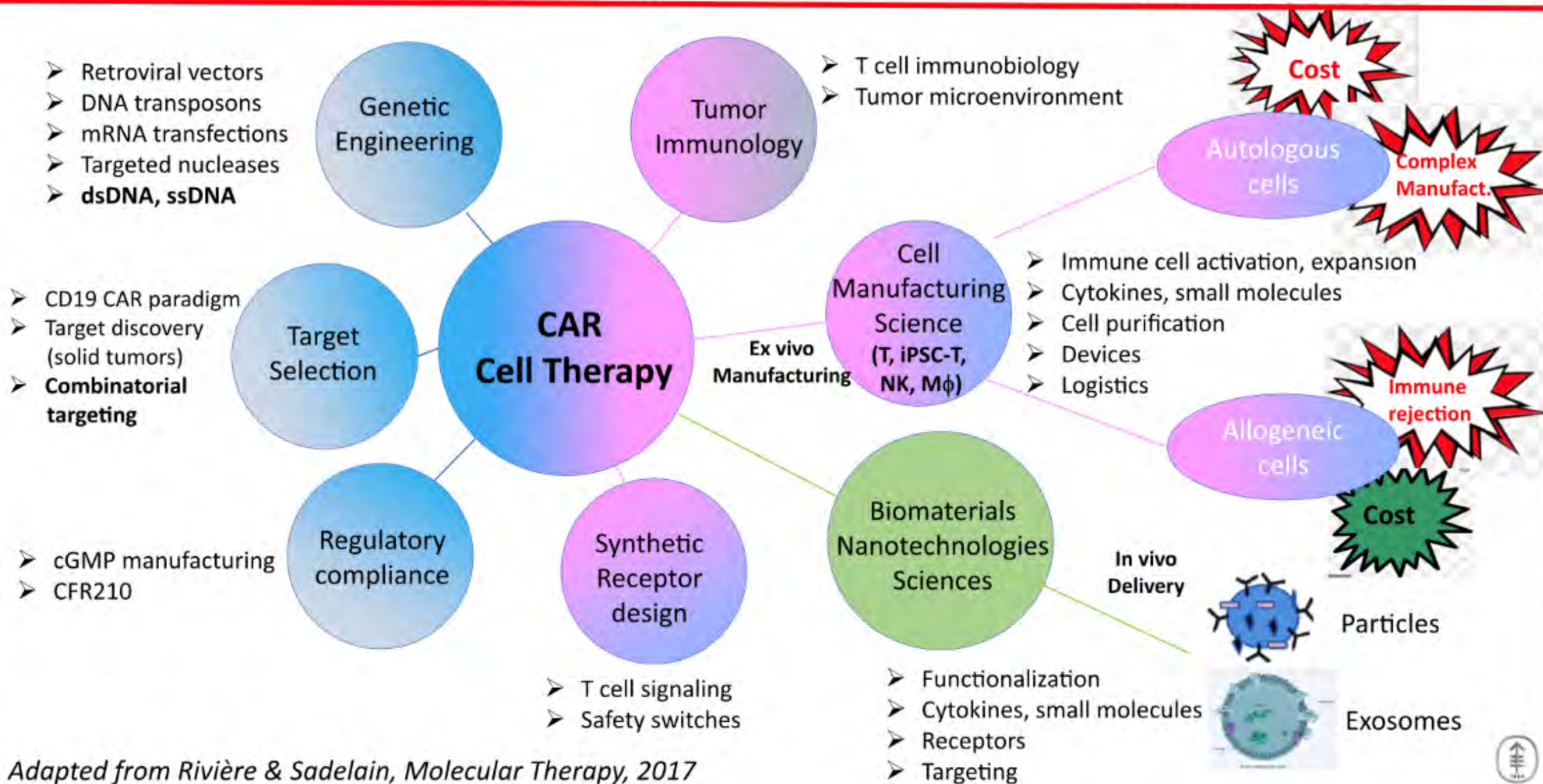
# Funnel approach drives the selection of the relevant features for the generation of a highly functional TCR T cell product



September 16<sup>th</sup> 2021

US **FDA** acceptance of investigational new drug application **NTLA5001**, CRISPR/Cas-engineered TCR-T cell candidate for **acute myeloid leukemia**

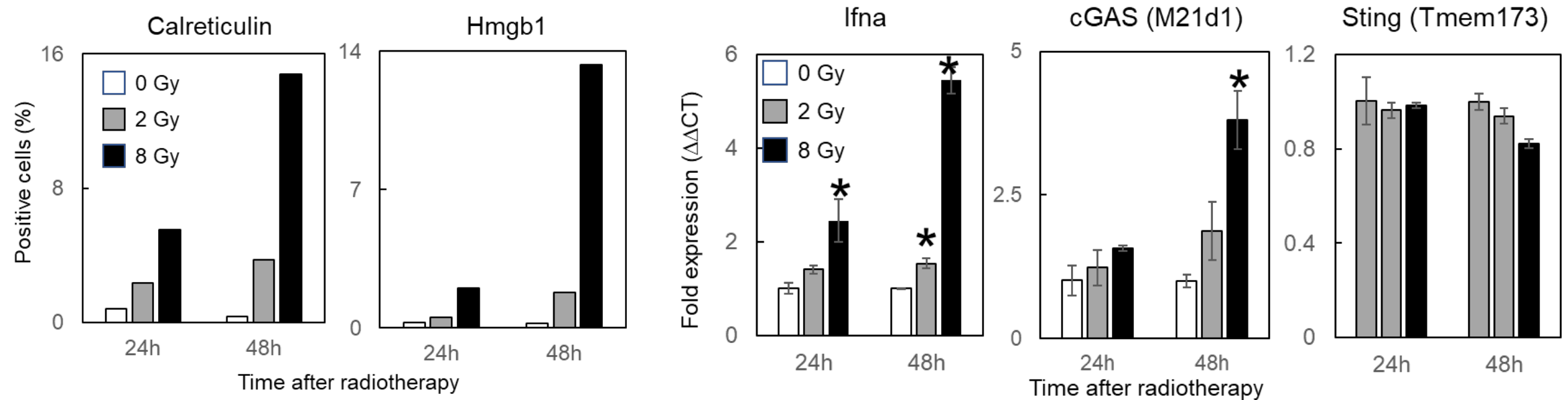
- Pipeline for the isolation of tumor-specific TCRs from healthy donors and cancer patients' circulating T cells
- Simultaneous editing of endogenous TCR  $\alpha$  and  $\beta$  chain genes using CRISPR/Cas9 technology (efficiency >90%)
- Transduction of T-cells with lentiviral vectors encoding WT1-specific TCRs (efficiency >95% of CD8<sup>+</sup> T cells)
- Targeted TCR KI in TRAC and simultaneous TRBC KO (efficiency 70-80%)
- TCR edited T-cells specifically and efficiently kill primary WT1<sup>+</sup> leukemic blasts in vitro and in vivo



Adapted from Rivière & Sadelain, *Molecular Therapy*, 2017

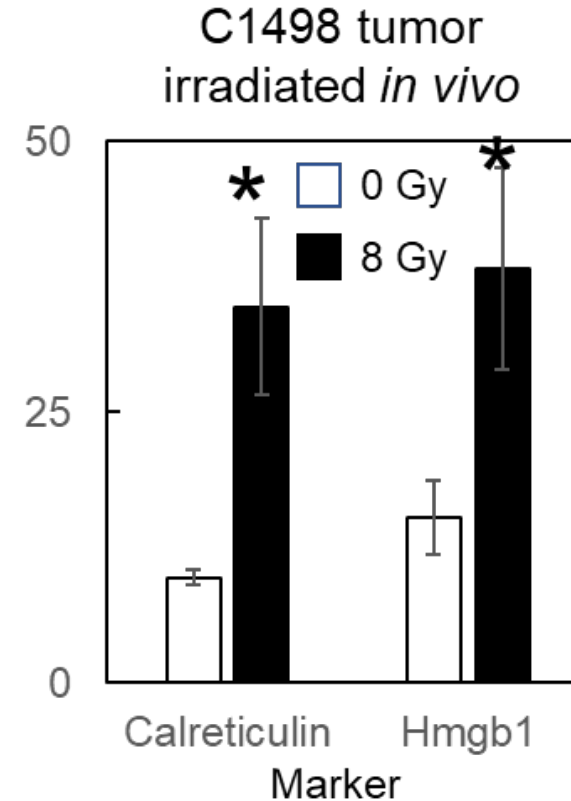
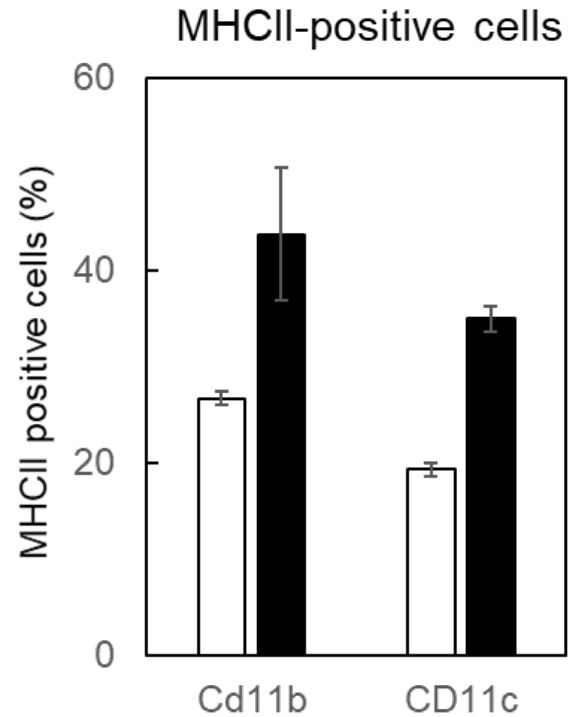
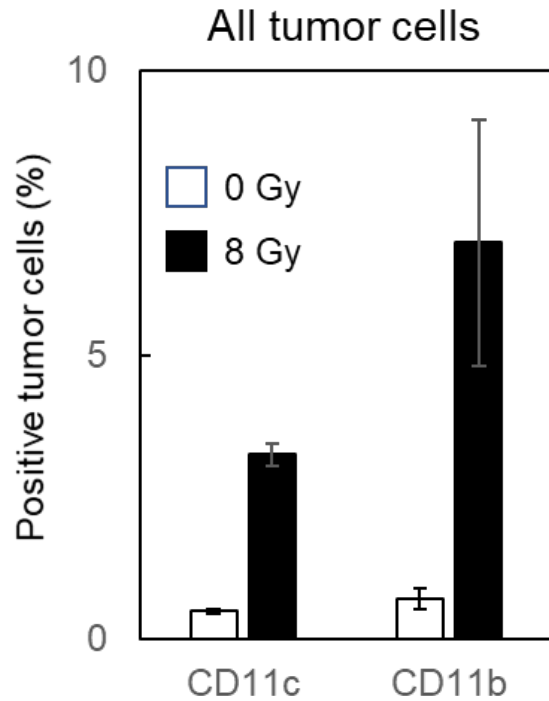


# Radiotherapy increases DAMP and cGAS-STING-IFN pathway expression *in vitro*



# Radiotherapy increases macrophage infiltration and DAMP expression *in vivo*

Tumor cells isolated irradiated C1498 tumors *in vivo*:







# ACT with engineered T cells

## Challenges and Opportunities

### Increasing function and persistence of T cells

*Bondanza et al, Blood 2006*  
*Kaneko et al, Blood, 2009*  
*Cieri et al, Blood, 2013*  
*Oliveira et al, Science Transl Med 2015*  
WO 2007017915  
WO 2021219758



Wendell A.L. & June C.H., *Cell*, 2016

### Redirecting T cell specificity (CAR & TCR)

*Casucci et al, Blood 2013*  
*Norelli et al, Nat Med 2018*  
*Greco et al, Sci Transl Med 2022*  
*Provasi, Genovese et al Nat Med 2012*  
*Mastaglio et al, Blood 2017*  
*Ruggiero et al, Sci Transl Med 2022*  
US P.A.N. 12/927,292  
Patent N. 9937207

### Remodeling of microenvironment Identifying suppression mechanisms (checkpoint inhibitors)

*Noviello, Manfredi et al Nat Comm 2019*  
*Toffalori et al, Nat Med 2019*

### Increasing the safety profile of T cells (suicide genes)

*Bonini et al, Science 1997*  
*Bonini et al, Nat. Med. 2003*  
*Recchia et al, PNAS 2006*  
*Cicero et al, Blood 2007*

- Radiating a tumor initiates cancer cell death through immunogenic cell death markers, and chemokines in the tumor microenvironment leading to among others maturation of dendritic cells and macrophages, which in turn activates tumor-specific CD 8+ T cells that infiltrate the tumor. Antigens released from dying cancer cells facilitate the cross-presentation of tumor antigens. While activating the adaptive and innate immune response, radiation can also stimulate the differentiation of T regulatory via TGF and interleukin 10, inhibiting activated T cells that promote tumor progression.
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- Now that we know that radiation holds promise for its ability to positively modulate the immune system let us look at the current clinical evidence and how to best incorporate it.
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